



## Research Article

# Analysis of Effect of Genotype x Environment Interaction on Rice Grain's Iron Content in Indonesia using Graphical GGE-Biplot Method

Suwarto<sup>1\*</sup> and Nasrullah<sup>2</sup>

<sup>1</sup>Laboratory of Plant Breeding, Faculty of Agriculture, Jenderal Soedirman University, Purwokerto, Indonesia; <sup>2</sup>Laboratory of Biometric, Faculty of Agriculture, Gadjah Mada University, Yogyakarta, Indonesia

\*Email: oryza\_07@yahoo.com

(Received:01 Feb 2011; Accepted:03Aug 2011)

### Abstract:

GGE-Biplot method was used to analyze data of iron (Fe) content of 10 rice genotypes evaluated at eight paddy fields during the wet and dry seasons of 2007 – 2008. The objective of this study was to determine the effect of genotype (G) and genotype x environment interaction (GE) on the iron content of 10 rice genotypes. Experiments were conducted using a randomized completely block design with three replications at each environment. Results indicated that environment (E), G and GE had significant effect on rice grain's iron content. Environment explained 74.43% of the total (G + E + GE) variation, whereas G and GE explained 5.60% and 19.67% of the total variation, respectively. The first two principal components (PC1 and PC2) were used to create a two-dimensional GGE-biplot, and these principal components explained 70.40% and 15.36% of the GGE sum of squares, respectively. Barumun genotype was desirable for its highest iron content, especially in term of ability and stability. Cilongok environment was the best representative of the overall environments and the most powerful environment to discriminate the genotypes.

**Key words:** Rice, iron content, genotype×environment interaction, GGE-biplot

### Introduction

Micronutrient trait expression and the extent of genotype×environment interactions across different environments largely determine screening, breeding, and testing methodologies used, and reflect trait heritability, genetic variation, and, hence, potential genetic gain. For the purpose of breeding for increased iron (Fe) content in rice (biofortification), the stability of Iron-dense traits of genotype across environment is inevitable (Suwanto and Nasrullah, 2011).

Crop performance, the observed phenotype, is a function of genotype (G), environment (E) and genotype×environment interaction (GE). GE is commonly observed as differential ranking of cultivar performances among locations or years. GE is said to occur when different cultivars or genotypes respond differently to diverse environments. Researchers have long been aware of the various implications of GE in breeding program. GE is important only when it is significant and causes significant change in genotypes' ranks in different environments (Crossa, 1997). GE has a negative

impact on heritability. A significant GE can seriously impair efforts to selecting superior genotypes for crop introductions and cultivar development programs (Yan and Racjan, 2002). Knowing the effect of GE, as well as the estimate of its magnitude relatives to the magnitude of G and E effects is very important for efficient breeding program. Therefore, understanding the structure and nature of GE is particularly useful to breeders as it help determine whether to develop cultivars for all environments or to develop specific cultivars for specific target environments (Bridges, 1989).

Data collected in multi-location trials are intrinsically complex, thus they should be analyzed and presented in a form of which it is easy to be understood, and interpreted with high accuracy. Many analysis methods have been developed to handle multi-location trials data, such as variant component, regression approach, Additive Main Effect and Multiplicative Interaction (AMMI) and most recently was Genotype and Genotype × Environment (GGE). Zobel *et al.* (1988) compared the traditional statistical analysis, analysis of variance (ANOVA), principle

component analysis (PCA) and linear regression, with AMMI analyses, and showed that traditional analyses were not always effective in analyzing multiple-environment trial (MET) data structure. ANOVA is an additive model that describes main effects effectively and determines if GE is a significant source of variation, but it does not provide insight into the patterns of genotypes or environments that give rise to the interaction. Principal component analysis is a multiplicative model that contains no sources of variation for additive G or E main effects and does not analyze the interactions effectively. Linear regression method uses E means, which are frequently a poor estimate of environments, such that the fitted lines in most cases account for a small fraction of the total GE.

Linear-bilinear models are useful tools for analysing MET data, and examining and interpreting genotype  $\times$  environment interaction (GEI) (Crossa and Cornelius, 1997). Useful linear-bilinear models, among others, are the AMMI model and the Sites Regression (SREG) model.

Biplots are useful for summarizing patterns of response that exist in the original data. Biplots graph scores of sites and genotypes of the first bilinear term against scores of sites and genotypes of the second bilinear term. A full description of the interpretation of the biplots of multiplicative models is given in Gower and Hand (1996). Briefly, the cultivar and site scores are represented as vectors in a two-dimensional space. The cultivar and site vectors are drawn from the origin (0, 0) to the end points determined by their scores. An angle less than  $90^\circ$  or larger than  $270^\circ$  between a cultivar vector and a site vector indicates that the cultivar had a positive response at that site. A negative cultivar response is indicated if the angle is between  $90^\circ$  and  $270^\circ$ . The cosine of the angle between two sites (or cultivars) approximates the phenotypic correlation of the two sites (or cultivars) with an angle of zero indicating a correlation of +1, an angle of  $90^\circ$  (or  $-90^\circ$ ) a correlation of 0, and an angle of  $180^\circ$  a correlation of -1.

A standard biplot of the SREG model was reported by Yan et al. (2000). He proposed to connect the scores of the furthest cultivars in the biplot such that they are at the corners of an external polygon and, for each side of the polygon, drawing a line segment perpendicular to that side that passes through the origin. These line segments subdivide the polygon into sectors involving different subsets of sites and cultivars. The genotype that is at the corner of one sector is the best performer in the sites included in

that sector. Sites located far away from the origin discriminate the cultivars more than those near the origin.

The GGE-biplot refers to genotype main effect (G) plus genotype by environment interaction (GE) which is the two sources of variation of the SREG model. The biplot from the SREG model shows that ideal genotypes should have large primary effects (high mean yield) and near zero secondary effects (more stable) and the ideal sites should have large primary effects (high power to discriminate cultivars) and small secondary effects. Such properties tend to occur if the primary effects of cultivars are highly correlated with the cultivar means (Yan et al., 2000; Crossa et al., 2001).

### Material and methods

Ten rice genotypes that represent the major genotypes grown in commercial fields of Central Java, Indonesia, were used in this study. They were Bahbutong(G1), Cimelati(G2), Fatmawati(G3), Barumun(G4), Aeksibundong(G5), IR64(G6), Sintanur(G7), Bengawan Solo(G8), Maligaya Special(G9) and Gilirang(G10). Yield potential of these genotypes ranges from high (Fatmawati) to good (other genotypes), milling quality ranges from excellent (Cimelati) to average (IR64), and maturity ranges from very early (Maligaya Special) to mid-season (Barumun). These rice genotypes were evaluated at four locations in Central Java, Indonesia, namely: Banyumas, Gombang, Cilongok and Baturaden during the wet and dry seasons of 2008. At each location, a randomized complete block design with three replications was used to arrange treatments. Each experimental plot was of size  $2 \times 10 \text{ m}^2$ . Grain yield was obtained from a harvested area of  $13.12 \text{ m}^2$  at the center of each plot.

Iron contents in rice grains were analyzed using 200 g of whole grain sample (non-polished rice). Analysis of rice grain's iron content was carried out at the Laboratory of Food Sciences, Faculty of Agriculture, University of Jenderal Soedirman, Indonesia. Soil analysis was carried out in all experimental sites to determine Fe,  $\text{NO}_3\text{-N}$ , P, K contents and soil pH.

The collected data were analyzed using SAS 9.1 program where genotype was the fixed factor and growing seasons and locations were treated as random factors. The effects of G, E and GE were analyzed using Graphical GGE Biplot following Yan et al. (2000). Principal Component Analysis was used to analyze the association between rice grains iron content (factor 1) and environmental factors that

included soil Fe content (factor 2), soil NO<sub>3</sub>-N (factor 3), soil P content (factor 4), soil K content (factor 5) and soil pH (factor 6). The first two principal components (PC1 and PC2) were used to create a two-dimensional GGE-biplot.

### Results and discussion

Soil iron content at the experimental locations ranged from high (Cilongok) to low (Gombong), and soil pH ranged from high (Gombong) to low (Cilongok) (Table 1). Rice grain iron content was highly varied across locations and seasons (Table 2). Analysis of variance showed that environment, genotype and genotype x environment interaction significantly influenced rice grain iron content. Environment explained 74.43% of total (G + E + GE) variation, whereas G and GE captured 5.60% and 19.67%, respectively. Based on two-dimensional GGE-biplot, PC1 and PC2 explained 70.40% and 15.36% of GGE sum of squares. Effects of G, E and their interaction were significant on rice grain iron content. The effect of GE was three times of the contribution of G (Table 3, 4). Rice grain iron content was unstable depending on location. Iron content in wheat grains was also highly unstable (Oury *et al.*, 2006; Joshi *et al.*, 2010).

Identification of the best genotype in each environment: Visualization of the "which-won-where" pattern of MET data is important for studying the possible existence of different environments in a region (Gauch and Zobel, 1997; Yan *et al.*, 2000; Yan *et al.*, 2001). The polygon view of a biplot is the best way to visualize the interaction pattern between genotypes and environments and to effectively interpret a biplot (Yan and Kang, 2003). The vertex genotypes in this investigation were Barumun (G4), Maligaya Sp.(G9), Fatmawati (G3) and IR 64 (G6). The vertex genotype for each sector is the one that gave the highest rice iron content for the environments that fall within that sector (Figure 1). Another important feature of Figure 1 is that it indicates environment grouping, which suggests the possible existence of different environments. Only two sectors of the four sectors contained environments and these were identified as the two groups of environment. The group of environments that share the same best genotype (s) (identified as being located at the corner of the polygon) is termed the mega-environment (Yan *et al.*, 2000; Yan and Hunt, 2002). Hence, Barumun (G4) had the highest rice grains iron content at Cilongok wet season (E4), Maligaya Sp. (G9) had the highest rice grains iron content at Banyumas wet season (E1) and Baturaden wet season (E2).

Mean of performances and stability genotypes: Yield performance and stability of genotypes were evaluated by an average environment coordination (AEC) method (Yan, 2001; Yan and Hunt, 2002; Yan 2002). In this method, an average environment is defined by the average PC1 and PC2 scores of all environments, represented by small circle (Figure 2). Genotypes with above average rice grain iron content means were Barumun (G4), IR64 (G6) and Maligaya Sp.(G9). A longer projection to the AEC ordinate, regardless of the direction, represents a greater tendency of the GEI of genotype, which means it is more variable and less stable across environments or vice versa. Barumun (G4) was more stable as well as high yielding, and it was the best genotype in terms of better rice grain iron content mostly at Cilongok wet season.

Result from AMMI analyses also indicated that Barumun genotype was the best genotype in terms of better rice grains iron content and it was more stable as well as high yielding (Suwanto and Nasrullah, 2011). Result from AMMI and GGE-biplot analyses indicated that Barumun genotype was the most stable as well as high rice grains iron content. Both analyses indicated that Barumun was the best genotype.

Characterizing the environments using an environment x factor -biplot: The environment vectors are drawn to facilitate visualization of the interrelationship among factors (Figure 3). When they are well represented by the biplot, the cosine of the angle between the vectors of two factors approximates the correlation coefficient between them. Rice grain iron content (factor 1) was closely positively associated with soil Fe (factor 2), but closely negatively with soil pH (factor 6). Rice grains iron content was not associated with soil N (factor 3), soil P (factor 4) and soil K (factor 5).

### Conclusions

The results indicated a significant effect of genotype by environment interaction on the expression of rice grain iron content. Genotype, environment and their interaction represented 5.60%, 74.42% and 19.67% of total variance, respectively. An environment situated with high soil Fe and low pH such as in Cilongok during wet season is the ideal condition to select genotype for Fe content. Rice grain iron content is positively correlated with soil Fe, but negatively correlated with soil pH. GGE model analysis is useful to determine the relative performance of a genotype in a specific environment, and comparison of performances of genotypes across environments, comparison of two genotypes in

different environments, and to identify high yielding varieties across environments, the best varieties and the best environment.

### Acknowledgement

The authors thank the financial support from the Ministry of National Education, Indonesia, Rector of Jenderal Soedirman University, Purwokerto - Indonesia and the Post Graduate Programs of Gadjah Mada University, Yogyakarta - Indonesia.

### References

- Bridge WC Jr. 1989. Analysis of plant breeding experiment with heterogeneous variance using mixed model equation. *In: Applications of Mixed Models in Agriculture and Related Dicine.* So. Coop. Ser. Bull. No. 343. Baton Rouge, LA: Louisiana Agricultural Experiment Station: 145-151.
- Crossa J. 1997. Statistical analyses of multilocation trials. *Adv. Agron.*, 44:55-85.
- Crossa, J, Cornelius P L, Yan W. 2001. Biplot of linier-bilinier models for stability crossover genotype×environment interaction. *Crop Sci.*, 42: 619-633.
- Gauch HG Jr and Zobel RW. 1997. Identifying mega-environments and targeting genotypes. *Crop Sci.*,37: 311-326.
- Joshi AK, Crossa J, Arun B, Chad R, Trethowan R, Vargas M, Ortiz-Monasterio I. 2010. Genotype×environment interaction for zinc and iron content of wheat grain in eastern Gangetic plains of India. *Field Crops Res.*, 116: 268-277.
- Oury FX, Leenhardt F, Remesy C, Chanliaud E, Duperrier B, Balfourier F, Charmet G. 2006. Genetic variability and stability of grain magnesium, zinc and iron content in bread wheat. *Eur J Agron.*, 25: 177-185.
- Suwarto and Nasrullah. 2011. Genotype × environment interaction for Iron concentration of rice in Central Java of Indonesia. *Rice Sci.*, 18 (1) : 75 - 78.
- Yan, W., L.A Hunt., Q. Sheng, and Z. Szlavnic. 2000. Cultivar evaluation and mega-environment investigation based on GGE biplot. *Crop. Sci.*, 40 : 507-605.
- Yan, W., P.L. Cornelius, J. Crossa, and L.A. Hunt. 2001. Two types of GGE biplot for analyzing multi-environment trial data. *Crop. Sci.*, 41 : 656 - 663.
- Yan, W. 2001. GGE biplot – a windows application for graphical analysis of multienvironment trial data and other types of two-way data. *Agron. J.*, 93 : 1111-1118.
- Yan, W. 2002. Singular value partitioning in biplot analysis of multienvironment trial data. *Agron. J.* 94 : 990 – 996
- Yan, W. and L.A. Hunt. 2002. Biplot analysis of diallel data. *Crop Sci.*, 42 : 21 – 30.
- Yan, W., and M.S. Kang. 2003. GGE biplot analysis : A graphical tool for breeders, geneticist, and agronomists. CRS PRESS. Boca Raton, FL.
- Yan, W., and Racjan, I. 2002. Biplot analysis of test sites and trait relations of soybean in Ontario. *Crop Sci.* 42 : 11-20
- Zobel RW, Wright MJ, Gauch HGJr. 1988. Statistical analysis of a yield trial. *Agron J.*, 80: 388-393.

**Table 1. Mean of Fe, NO<sub>3</sub>-N, P, K contents in the soil and soil pH at the experimental locations in Central Java, Indonesia**

Parameter	Banyumas		Baturaden		Gombong		Cilongok	
	Wet season	Dry season						
	(E1)	(E5)	(E2)	(E6)	(E3)	(E7)	(E4)	(E8)
Fe (g/kg)	53.30	50.80	52.60	51.40	25.63	24.42	72.23	70.43
NO <sub>3</sub> -N(%)	0.12	0.14	0.17	0.15	0.13	0.11	0.03	0.03
P (mg/kg)	583.29	588.43	794.52	799.42	466.52	462.35	287.61	288.32
K (mg/kg)	345.12	332.52	485.48	480.83	327.78	324.68	227.60	221.20
pH	4.43	4.02	5.68	6.02	7.51	7.88	4.44	4.04

**Table 2. Rice grain iron content of 10 evaluated genotypes at eight environments in Central Java, Indonesia**

Genotype	Banyumas		Baturaden		Gombong		Cilongok (E4)		Average
	(E1)	(E5)	(E2)	(E6)	(E3)	(E7)	(E8)		
Bahbutong(G1)	20.60	6.76	14.72	9.68	24.80	10.51	46.26	20.51	19.22
Cimelati(G2)	18.04	9.27	16.93	13.52	16.55	16.40	48.20	23.44	20.29
Fatmawati(G3)	15.08	8.72	13.52	13.29	14.39	9.31	25.08	23.08	15.31
Barumun(G4)	18.63	8.25	30.70	16.67	19.96	16.46	77.70	28.87	27.15
Aeksibundong(G5)	20.58	9.70	20.85	9.70	12.34	10.92	45.50	20.14	18.71
IR 64(G6)	21.58	7.73	13.21	14.27	9.42	16.14	64.05	24.86	21.41
Sintanur(G7)	22.04	8.89	13.49	12.35	20.17	8.37	45.49	21.48	19.03
Bengawan solo(G8)	22.55	10.52	16.74	11.77	13.29	10.77	53.34	23.75	20.34
Maligaya Special(G9)	29.05	11.85	32.49	14.18	15.93	18.97	35.03	23.18	22.58
Gilirang(G10)	29.98	8.07	14.25	13.35	9.30	10.45	29.00	23.55	17.25
Average environment	21.81	8.98	18.69	12.88	15.62	12.83	46.96	23.29	

**Table 3. Analysis of variance of the effects of genotype, environment, genotype×environment (GE) on rice grains iron content**

Sources of variation	df	SS	MS	F value	P value	Model	SS(%)*
Environment	3	29569.269	4224.181	5577.70	<.0001	Random	74.42
Replication (Env)	16	12.098	0.756	1.00	0.5502		
Genotype	9	2222.953	246.994	326.14	<.0001	Fixed	5.60
Genotype x Env	63	7817.099	124.081	163.84	<.0001	Random	19.67
Residual	144	109.056	0.757				
Total	239	39730.477					

\* percentage from total SSG, E dan GE

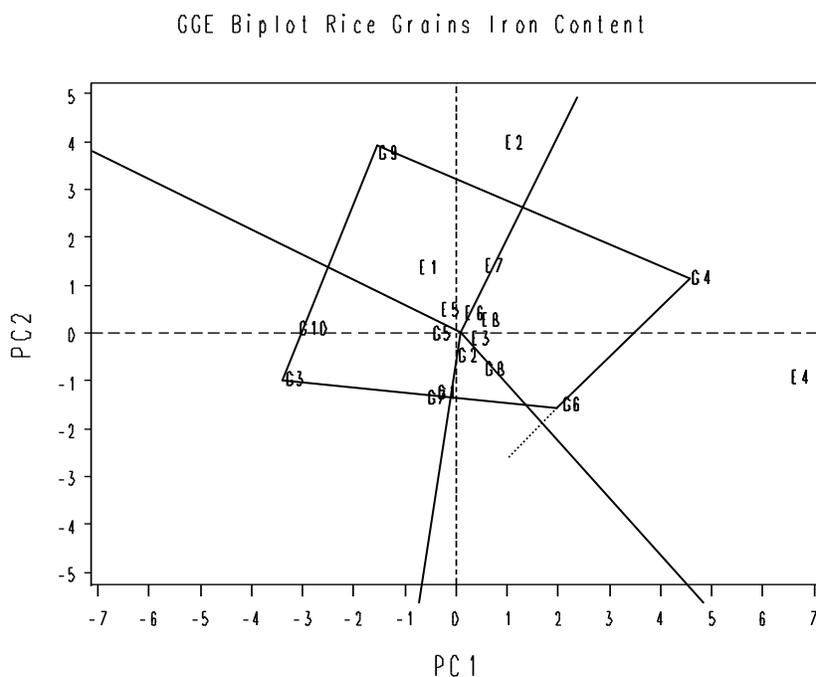
**Table 4. GGE biplot analysis of rice grain iron content of 10 evaluated genotypes at eight environments in Central Java, Indonesia**

Principal Component	Singular Values	Eigen Values	Total Eigen Values (%)	Cumulative (%)
PC1	84,07	7068,15	70,40	70,40
PC2	39,28	1542,76	15,36	85,76
PC3	27,69	767,13	7,64	93,40
Residu		662,00	6,60	100,00
Total		10040,04*		

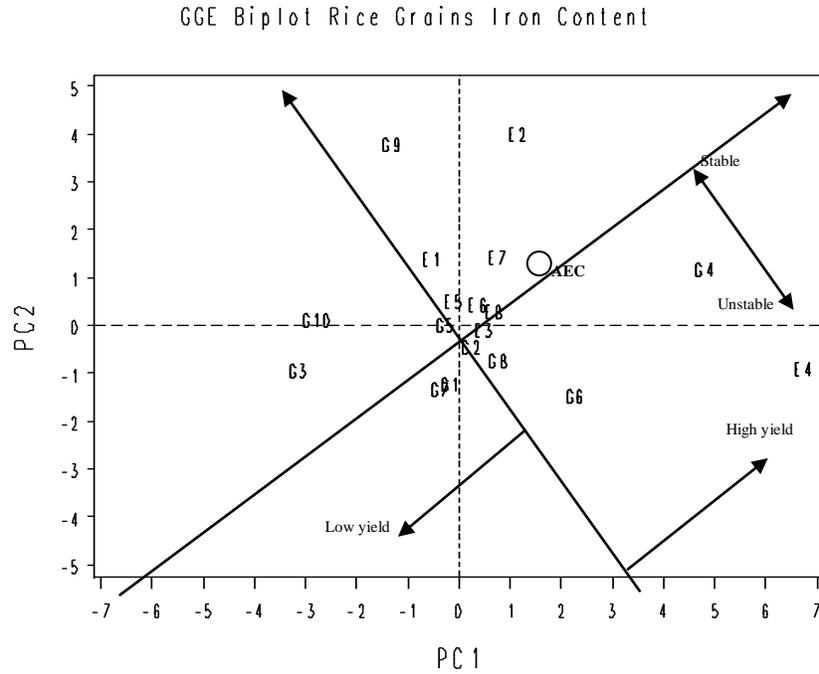
\* Total eigen values = total SS of Genotype + (Genotype x Environment)

**Table 5. Eigenvector 10 evaluated genotypes and the eight environments**

Factor	Code	PC1	PC2
Genotype	G1	-0.18049	-1.10891
Genotype	G2	0.24378	-0.34297
Genotype	G3	-3.15347	-0.82573
Genotype	G4	4.80065	1.27613
Genotype	G5	-0.27119	0.12212
Genotype	G6	2.27146	-1.36373
Genotype	G7	-0.38284	-1.21591
Genotype	G8	0.77021	-0.64479
Genotype	G9	-1.31202	3.89987
Genotype	G10	-2.78609	0.20390
Environment	E1	-0.55038	1.47425
Environment	E2	1.15301	4.09254
Environment	E3	0.48891	-0.00062
Environment	E4	6.74489	-0.80888
Environment	E5	-0.10878	0.59871
Environment	E6	0.37023	0.53132
Environment	E7	0.75262	1.51376
Environment	E8	0.67748	0.41020

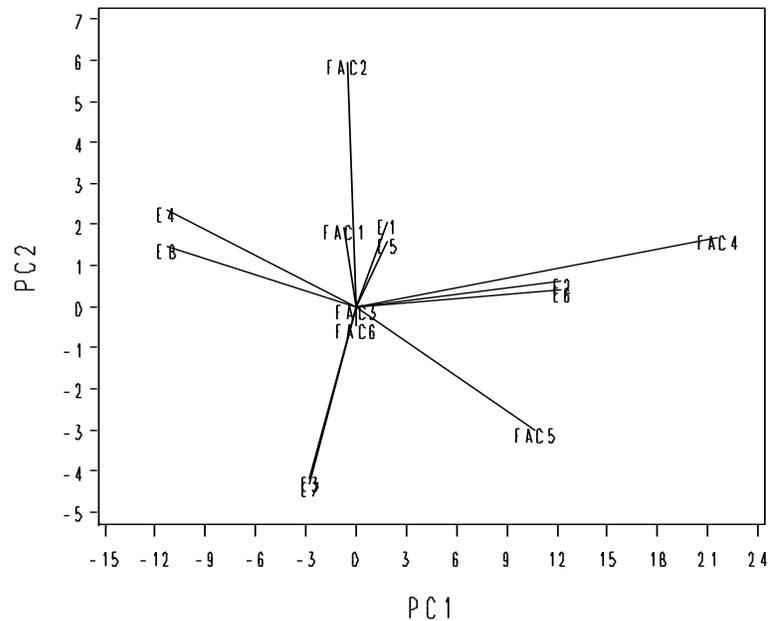


**Figure 1. Genotype plus genotype x environment (GGE) biplot of the environment and their winning genotypes**



**Figure 2. Average environment coordination (AEC) views of the GGE-biplot for the means performance and stability of genotypes**

Graphic-Biplot Rice Grains Iron content-Environment Factors



**Figure 3. Vector view of an environment x factor biplot summarizing the interrelationship among rice grain iron content (Fac 1) and soil factors. PC1 and PC2 are first and second principle components, respectively. Fac 2, soil iron content; Fac 3, soil NO<sub>3</sub>-N content; Fac 4, soil P content; Fac 5, soil K content; Fac 6, soil pH.**