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### **Research Article**

### Study on blackgram (*Vigna mungo* (L.) Hepper) genotypes for yield stability and Mungbean Yellow Mosaic Virus (MYMV) resistance

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#### Abstract

A total of 50 genotypes were evaluated during *Kharif 2022* in three different locations in Tamilnadu, India. The data on seed yield per plant was subjected to AMMI and GGE biplot analysis for stability study. The IPCA values explained 100% and 98.8% variation in AMMI and GGE biplot respectively. Results from both analyses inferred that the genotypes *viz.*, Mash 114, Mash 1008, VBN (Bg) 4 and VBN 10 were stable across the environments with high mean seed yield per plant. Hence these varieties can be utilized for cultivation in varied environments of Tamilnadu. Besides, the genotypes; MASH 1008, MASH 114, VBN (Bg) 4, VBN 8, VBN 10 and VBN 11 were found to be resistant to MYMV based on the field screening at National Pulses Research Centre, Vamban, Pudukkottai.

Keywords: Blackgram, Stability, AMMI, GGE Biplot, MYMV, Field Screening

#### INTRODUCTION

Blackgram [*Vigna mungo* (L.) Hepper] is an important short-duration pulse crop which is predominantly grown in South Asian regions during various seasons in rainfedbased agroecosystems (Kaewwongwal *et al.* 2015). It is packed with nutrients and forms a major component of a vegetarian diet because it contains about 24 - 26% protein in addition to other vital vitamins and minerals (USDA National Nutrient Database, 2018). India is the largest producer and consumer of blackgram (Vinothini and Kaleeswari 2022). Blackgram is grown in all seasons like *kharif, rabi,* summer irrigated and rice fallow situations. If a variety is suitable for all situations, it would be a gift to the farming community to raise the crop throughout the year without seasonal boundaries. To identify genotypes with predictable performance that are responsive to environmental variations, it is essential to analyze the adaptability and stability of each genotype to minimize the effects of the GE interaction. Even though numerous superior blackgram cultivars have been cultivated, many of them have primarily fluctuated in terms of yield performance in various regions due to significant GEI. Due to the unreliable performance of genotypes across environments because of GEI, the breeding programme ends up being increasingly challenging (Ebdon & Gauch, 2002). The AMMI model is flexible and effective for comprehending GEI. It shows GEI patterns graphically and encompasses both the additive and multiplicative components of the two-way data structure (Mukherjee *et al.*, 2013). Since, it explains more G+GE than AMMI, the GGE biplot is preferable in terms of mega environment analysis and genotype evaluation. The most widely applied statistical techniques for analyzing the results of multi-environment trials are AMMI and GGE biplot (Gauch 2006; Zhou *et al.*, 2011).

Mungbean Yellow Mosaic is one of the notorious diseases that impair blackgram productivity in most South Asian nations including India, Sri Lanka, Pakistan, and Bangladesh. This disease results in 50% yield reduction in blackgram (Dikshit *et al.*, 2020). It is transmitted by the whitefly, *Bemisia tabaci* (Nene, 1972), which is manageable with prompt insecticide administration. However, the inability and high expense of pesticides make it difficult to effectively control the disease. This research was carried out to gain insight into the nature and magnitude of genotype x environment interaction utilizing AMMI and GGE biplot analysis thus, identifying the stable performer among the genotypes in natural epiphytotic conditions during *kharif* season in hotspot region.

#### MATERIALS AND METHODS

Stability analysis for Seed yield: The experiment was carried out in three locations during Kharif 2022 at Agricultural College and Research Institute, Madurai (E1), National Pulses Research Institute, Vamban (E2) and a farmer's field at Vizhudhudayan, Ariyalur district (E3). The details of the environments are presented in Table 1. A total of 50 diverse blackgram genotypes were collected from various sources such as National Pulses Research Centre, Vamban; Indian Institute of Pulses Research, Kanpur; Agricultural Research Station, Lam and Tamil Nadu Agricultural University, Coimbatore and were evaluated in a randomized complete block design with two replications in each environment. The data on seed yield per plant (g) was collected from 10 plants per replication and subjected to stability analysis to identify the stable genotype. The genotype x environment (GxE) interaction was studied as per the AMMI model (Zobel et al., 1988) and GGE biplot (Yan, 1999 and Yan et al., 2001). All the analyses were performed using the software PBtools (ver 1.3), International Rice Research Institute, Philippines.

MYMV Screening: The 50 blackgram genotypes were screened for mungbean yellow mosaic virus (MYMV) at National Pulses Research Centre, Vamban in Kharif 2022 which is a hotspot for MYMV. The susceptible check, CO 5 was raised in every 10<sup>th</sup> row to ensure the necessary spread of MYMV as per the infector row technique. All the recommended packages of practices were carried out except for insecticide spray to encourage the building of the whitefly population in the field. The scoring was performed on the 45<sup>th</sup> day and the 60<sup>th</sup> day after sowing when the susceptible check, CO 5 showed a maximum scale of 9 for MYMV. The arbitrary score scale of 0-9, proposed by Singh et al. (1995) which indicates 1 free (F), 2 highly resistant (HR), 3- resistant (R), 4- moderately resistant (MR), 5-moderately susceptible (MS), 6susceptible (S), 7- susceptible (S), 8- highly susceptible (HS) and 9- highly susceptible (HS), was used for scoring.

#### **RESULTS AND DISCUSSION**

Analysis of variance for each environment revealed that the genotypes were significant in all three environments representing a greater magnitude of genetic variation for the trait, seed yield per plant (g). Further, pooled analysis of variance exhibited significant GxE interaction for seed yield per plant (Table 2), hence, stability analysis was performed. Genotypes typically showed variations in their performance in response to different environmental conditions. Stability analysis is an efficient tool used by the plant breeders to comprehend, how a genotype responds to its surroundings. To improve seed yield, stable-performing genotypes must be developed. When developing varieties, the relationship between genotype and environment is crucial. Adaptability and character expression may vary according to the environmental condition, leading to developmental variation in their performances across different environments. Therefore, developing varieties with stable performance across different locations of district and state will be rewarding to improve the overall productivity and production of blackgram.

The AMMI is a hybrid model that includes the two-way data structure's additive and multiplicative parts. The AMMI analysis is an invaluable tool for graphically defining GxE outlines. The biplot display of PCA scores stacked one on top of the other provides a graphic representation and interpretation of the GxE interaction. The combination of biplot presentation and genotypic stability statistics permits the grouping of genotypes based on resemblance

Code	Environment	Agroecological zone	Rainfall (mm)	Soil type	Latitude	Longitude	Altitude(m)
E1	Madurai	Southern	855	Sandy Clay loam	9°58' N	78°12' E	134
E2	Vamban	Southern	881	Red laterite	10°22' N	78°54' E	93
E3	Vizhudhudayan	North Eastern	1050	Red loam	11°24' N	79°20' E	45

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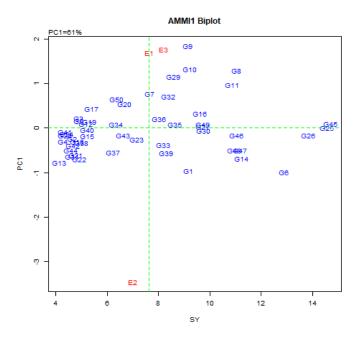
Character	Df		Seed yield per plant (g)
Genotypes (G)	49		26.62**
Environment	2		18.03**
Genotype x Environment	98		5.40**
IPCA 1	50	MSS	12.91**
		% Explained	61.00
IPCA 2	48	MSS	8.59**
		% Explained	39.00
MS due to Pooled error	147		0.49

Table 2. ANOVA for stability (AMMI) analysis for seed yield per plant (g)

in performance through diverse locations (Mukherjee *et al.*, 2013). The analysis of variance for AMMI showed that the interaction principal component axes *viz.*, IPCA 1 and IPCA 2 were significant for the trait, seed yield per plant (g). The contribution of IPCA 1 and IPCA 2 towards interaction was observed to be 61 and 39% respectively. This revealed that both the components together fully explained the GxE interaction for seed yield per plant (g). Similar results were reported by Dhasaradhan *et al.* (2021) and Sridhar *et al.* (2023).

A stable genotype is one whose performance is constant despite variations in the environment (Karimizadeh *et al.*, 2013). Among the genotypes; G6, G8, G10, G11, G14, G16, G25, G26, G27, G30, G45, G46, G47, G48 and G49 had significantly higher mean seed yield per plant than the grand mean. However, the genotypes, Mash 1008 (25), Mash 114 (G26), VBN (Bg) 4 (G45), VBN 10 (G46) and VBN 8 (G48) exhibited higher mean and IPCA scores nearer to '0' indicating that they are stable genotypes with less environmental interaction (**Table 3** and **Fig. 1**),

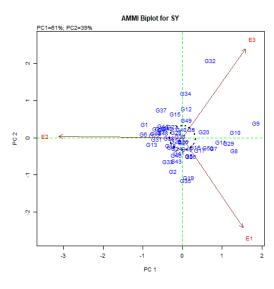
which infer that they are suitable for all environments. An environment with the longest spoke is considered more interactive. E2 has the longest spoke of the three environments, implying that it is more interactive than the other two. The AMMI 2 biplot (Fig. 2) provides plenty of information about the interaction component but fails to demonstrate the additive main effects. When it comes to AMMI 2 biplots, a genotype is thought to be more stable if it is situated close to the biplot's origin, or in its centre. A genotype with a high mean yield coupled with a high IPCA value indicates the specific adaptability to environments, hence the genotypes; VBG 19-033 (G1), IPU 19-51 (G6), DPU-552 (G13) and KKB 19-005 (G31) can be recommended for cultivation in the specific environment, E2. Other high-yielding genotypes like TBU 236-6 (G9), DPU-238 (G12), CO BG 21-04 (G32) and NIC 15-26 (G34), implying that they constructively interact with the environment and can therefore be encouraged for cultivation in the favorable environment, E3. Based on the AMMI biplots 1 and 2, the genotypes; Mash 1008, Mash 114, VBN (Bg) 4 and VBN 10 were identified as stable

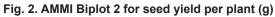




Code	Genotype	Seed yield per plant (g)			Code	Genotype	Seed yield per plant (g)		
		Mean	IPCA1	IPCA2			Mean	IPCA1	IPCA2
G1	VBG 19-033	9.17	-0.97	0.36	G29	VBG 18-099	8.58	1.16	-0.14
G2	SVU 6	4.64	-0.24	-0.90	G30	KKB 19-003	9.75	-0.05	0.03
G3	LBG 787	4.88	0.22	0.22	G31	KKB 19-005	4.64	-0.65	-0.04
G4	1PU 13-6	4.27	-0.30	-0.26	G32	CO BG 21-04	8.38	0.71	2.08
G5	VBG 17-021	4.89	0.16	-0.48	G33	N-1	8.20	-0.37	-0.64
G6	IPU 19-51	12.88	-0.99	0.10	G34	NIC 15-26	6.35	0.08	1.21
G7	IPU 18-7	7.67	0.78	-0.29	G35	PBD-103	8.64	0.09	-1.16
G8	MBG 1080	11.04	1.30	-0.34	G36	PDU-31	8.01	0.21	-0.50
G9	TBU 236-6	9.14	1.85	0.39	G37	PLU 446	6.24	-0.54	0.76
G10	C0 5	9.22	1.33	0.14	G38	PLU-1079	5.00	-0.34	-0.01
G11	CO BG 45	10.87	0.97	-0.11	G39	PLU-294	8.29	-0.56	0.24
G12	DPU-238	5.18	0.10	0.78	G40	PLU-608	5.23	-0.04	0.22
G13	DPU-552	4.13	-0.77	-0.17	G41	RU 9-195	4.35	-0.09	-0.41
G14	DPU-6	11.24	-0.68	0.12	G42	STU-22-88	4.66	-0.39	0.25
G15	DPU-99-228	5.22	-0.19	0.65	G43	CO 7	6.62	-0.16	-0.63
G16	IPU 99-224	9.62	0.33	-0.26	G44	VBG 06-019	4.58	-0.50	0.32
G17	KKG-0-3	5.40	0.43	-0.34	G45	VBN (Bg) 4	14.71	0.10	-0.30
G18	KU 9-157	4.84	-0.30	-0.21	G46	VBN 10	11.05	-0.16	-0.48
G19	KU-9-152	5.31	0.15	-1.08	G47	VBN 11	11.18	-0.50	0.22
G20	KU-9-158	6.69	0.55	0.16	G48	VBN 8	12.96	-0.50	0.13
G21	L 13-10	4.77	-0.61	0.24	G49	VBN 9	10.73	0.09	0.48
G22	L-401	4.92	-0.71	0.07	G50	VH 86-47	6.38	0.65	-0.27
G23	L-64	7.16	-0.26	0.30	E1	Madurai	8.22	1.69	-2.70
G24	LBG 17	4.42	-0.14	-0.29	E2	Vamban	7.01	-3.46	0.04
G25	MASH 1008	14.58	0.01	-0.10	E3	Vizhudhudayan	7.66	1.77	2.66
G26	MASH 114	13.85	-0.17	-0.08		Mean	7.63		
G27	MDU 1	9.72	0.03	-0.13		SE	1.49		
G28	VBG 19-010	4.36	-0.16	0.15		CD (5%)	1.87		

Table 3. Mean and IPCA scores of the genotypes and environments for seed yield per plant (g)





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genotypes with high yield potential. The environment which can separate the genotypes should have a high IPCA1 and a low IPCA2. Among the environments, Madurai (E1) had the highest mean for seed yield per plant making it the most favorable environment followed by Vizhudhudayan (E3) and Vamban (E2). Vamban (E2) is the most interactive and discriminative environment so it is a good test environment to cull out the unstable genotypes.

Numerous studies showed that, the environment influences more with the highest magnitude, whereas the main genotype effect and GEI (Genotype Environment Interaction) had a lesser effect in explaining variations. Since, environment is an uncontrollable component, GGE biplot graphically visualizes G plus GE of a MET in a way that makes genotype evaluation and mega environment identification easier. GGE biplot graphically partitions the complex GEI into two distinct principal components (Yan et al., 2000; Yan et al., 2007). The GGE biplot effectively captured 98.8% of variation via PC1 (71.8%) and PC2 (27.0%) respectively. This demonstrated that the biplot constructed by the first principal component scores of genotypes and environments against the respective scores of the second principal component adequately represented the environment-centered data. These findings were in harmony with the earlier reports by Jeberson et al. (2022) and Parmar et al. (2022) in blackgram, wherein they also observed 90% of the total GGE variation in the first two principal components.

The Average Environment Axis (AEA) represents the average coordinate of all the environments being studied. The environment, E1 and AEA bring about an acute angle making it essentially representative of all test environments, whereas E2 is the least representative

environment (Fig. 3). E1 is a suitable test environment for choosing genotypes that are generally acclimated. Since E2 is the least representative and most discriminative, it can be used to rule out unstable genotypes. E1 and E3 together make up a single mega environment, whereas, E2 forms a single distinct mega environment. The Average Environment Coordination Abscissa (AEA), which has just a single arrow, implies that the mean seed yield is escalating. The AEC coordinate with two arrows on it shows the most variation in either direction. With this perspective, the genotypes, G6 (IPU 13-6), G14 (DPU-6), G17 (KKG-0-3), G25 (Mash 1008), G26 (Mash 114), G45 (VBN (Bg) 4) and G46 (VBN 10) were ideal with high mean and less interactive with the environment (Fig. 4). The "which won-where" biplot as a polygon (Fig. 5) is the best way to interpret the interaction patterns between genotypes and environments and give comprehensive information. The polygon is drawn by joining the farthest genotypes from the biplot origin in such a way that all the other genotypes are within the polygon. The perpendicular lines connecting the sides of the polygon generate parts of genotypes and environments (Hernandez and Crossa 2000). The genotypes on the polygon's vertices indicate which genotypes perform more effectively in specific environments. The winning genotypes are situated on the vertices of each sector. E1 and E3 are grouped under a single mega environment while, E2 in a separate mega environment with respective winning genotypes at the vertex. The genotype, G1 (VBG 19-033) and G6 (IPU 19-51) are the winning genotypes in the mega environment comprising of E2, whereas, G25 (Mash 1008), G26 (Mash 114) and G45 (VBN (Bg) 4) are winning genotypes in mega environment comprising E1 and E3. Hence, for different mega environments, different genotypes can be identified and suggested for cultivation in the respective locations.

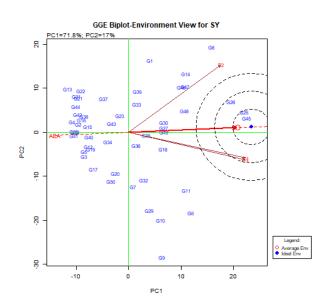


Fig. 3. GGE Biplot- Environment view for seed yield per plant (g)



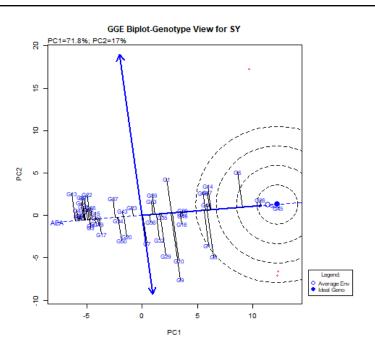


Fig. 4. GGE Biplot- Genotype view for seed yield per plant (g)

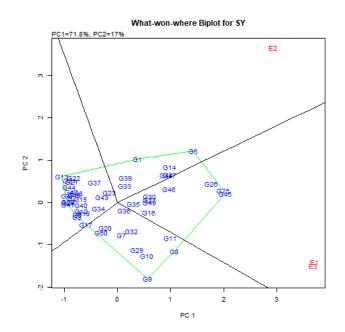


Fig. 5. GGE Biplot- Which won where plot for seed yield per plant (g)

Field screening for MYMV disease resistance:Yellow Mosaic Virus was initially discovered in 1960 in Dolichos (Capoor and Varma, 1950) and in mungbean (Nariani, 1960). It is one of the most destructive viral diseases that lowers productivity of the crop. The reduction in yield is largely due to susceptibility to MYMV (Alam *et al.*, 2014). The phenotypic scoring for MYMV was performed according to Singh *et al.* (1995).

A total of 50 genotypes were categorized into 6 groups based on the MYMV score. The genotypes *viz.*, MASH 1008, MASH 114, VBG 18-099, VBG 06-019, VBN (Bg) 4, VBN 10 and VBN 8 were highly resistant whereas, the genotypes; VBG 19-033, SVU 6, DPU-238, DPU-552, DPU-6, DPU-99-228, L 13-10, L-401, KKB 19-005, PDU-31, PLU-294, PLU-608, VBN 11and VBN 9 were resistant to MYMV (**Table 4**). Similarly, Mash 114 has been

S. No.	Mymv Score	Disease Reaction	Number Of Genotypes	Genotypes
1.	2	Highly Resistant (HR)	7	MASH 1008, MASH 114, VBG 18-099, VBG 06-019, VBN (Bg) 4, VBN 10, VBN 8
2.	3	Resistant (R)	14	VBG 19-033, SVU 6, DPU-238, DPU-552, DPU-6, DPU-99- 228, L 13-10, L-401, , PDU-31, KKB 19-005, PLU-294, PLU- 608, VBN 11, VBN 9
3.	4	Moderately Resistant (MR)	9	1PU 13-6, VBG 17-021, IPU 19-51, KU 9-157, KU-9-152, PLU 446, PLU-1079, RU 9-195, CO 7
4.	5	Moderately Susceptible (MS)	7	LBG 787, IPU 18-7, IPU 99-224, KKG-0-3, L-64, VBG 19- 010, KKB 19-003
5.	6	Susceptible (S)	3	N-1, PBD-103, STU-22-88
6.	7	Susceptible (S)	4	VH 86-47, MBG 1080, TBU 236-6, CO BG 45
7.	8	Highly Susceptible (HS)	3	CO BG 21-04, NIC 15-26, MDU 1
8.	9	Highly Susceptible (HS)	3	C 5, KU-9-158, LBG 17

Table 4. MYMV disease reaction of the 50 genotypes based on Singh et al. (1995)

reported to be highly resistant to MYMV by Gill (2018). Nine genotypes *viz.*, 1PU 13-6, VBG 17-021, IPU 19-51, KU 9-157, KU-9-152, PLU 446, PLU-1079, RU 9-195 and CO 7 were identified to be moderately resistant while the other genotypes fall under susceptible category. Hence, the varieties that were highly resistant to MYMV can be utilized as resistant donors in breeding programs.

With regard to the graphical models of AMMI and GGE, the genotypes; Mash 1008, Mash 114, VBN (Bg) 4 and VBN 10 were found to be stable and also had high yield potential. The stable genotypes identified from this study can be used for future breeding programs. High-yielding genotypes for specific environments may be investigated in larger plots and can be used for recombination breeding to obtain ideal segregants for sustainable blackgram production. Besides, the genotypes; MASH 1008, MASH 114, VBG 18-099, VBG 06-019, VBN (Bg) 4, VBN 10 and VBN 8 were highly resistant to mungbean yellow mosaic virus. So, in combination of yield stability and MYMV resistance, the varieties, Mash 1008, Mash 114, VBN (Bg) 4 and VBN 10 were adjudged as best and can be recommended for cultivation for all three environments and also can be used in hybridization to evolve superior, stable and MYMV resistant blackgram genotypes.

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#### REFERENCES

Capoor, S.P. and Varma, P.M. 1950. A new virus disease of Dolichos lablab. *Current Science*, **19 (8).** 

- Dhasarathan, M., Geetha, S., Karthikeyan, A., Sassikumar, D. and Meenakshiganesan, N. 2021. Development of novel blackgram (*Vigna mungo* (L.) Hepper) mutants and deciphering genotype× environment interaction for yield-related traits of mutants. *Agronomy*, **11(7)**: 1287. [Cross Ref]
- Dikshit, H. K., Mishra, G. P., Somta, P., Shwe, T., Alam, A. K. M. M., Bains, T. S., et al., 2020. Classical genetics and traditional breeding in mungbean. in *The Mungbean Genome, Compendium of Plant Genomes* ed. Nair, R. M., et al (Switzerland AG: © Springer Nature), 43–54. [Cross Ref]
- Ebdon, J.S. and Gauch Jr, H.G., 2002. Additive main effect and multiplicative interaction analysis of national turfgrass performance trials: I. Interpretation of genotype× environment interaction. *Crop science*, **42(2)**: 489-496. [Cross Ref]
- Gauch Jr, H.G. 2006. Statistical analysis of yield trials by AMMI and GGE. *Crop science*, **46(4**): 1488-1500. [Cross Ref]
- Gill, R.K. 2018. Identification of molecular marker linked to mungbean yellow mosaic virus (MYMV) resistance in Vigna radiata (L.) Wilczek. *Electronic Journal of Plant Breeding*, 9(3): 839-845. [Cross Ref]
- Hernandez M. V. and Crossa J. 2000.The AMMI analysis and graphing the biplot. Biometrics and Statistics Unit, CIMMYT, Mexico.
- Jeberson, M.S., Parihar, A.K., Shashidhar, K.S., Dev, J., Dar, S.A. and Gupta, S., 2022. Selection of suitable genotypes of urdbean (*Vigna mungo* L.) for targeted environments of hilly terrains of India using GGE biplot and ammi analysis. *Legume Research-An International Journal*, **45(6)**: 669-675.
- Karimizadeh, R., Mohammadi, M., Sabaghni, N., Mahmoodi, A.A., Roustami, B., Seyyedi, F. and Akbari, F.,

2013. GGE biplot analysis of yield stability in multienvironment trials of lentil genotypes under rainfed condition. *Notulae Scientia Biologicae*, **5(2):** 256-262. [Cross Ref]

- Kaewwongwal, A., Kongjaimun, A., Somta, P., Chankaew, S., Yimram, T. and Srinives, P. 2015. Genetic diversity of the blackgram [*Vigna mungo* (L.) Hepper] gene pool as revealed by SSR markers. *Breed. Sci.*, 65: 127–137. [Cross Ref]
- Mukherjee, A.K., Mohapatra, N.K., Bose, L.K., Jambhulkar, N.N. and Nayak, P., 2013. Additive main effects and multiplicative interaction (AMMI) analysis of GxE interactions in rice-blast pathosystem to identify stable resistant genotypes. *African Journal* of *Biotechnology*, 8(44): 5492-5507.
- Nariani, T.K. 1960. Yellow mosaic of Mung (*Phaseolus aureus* L.). *Indian Phytopathology*, **13 (1).**
- Nene, Y.L. 1972. A survey of viral diseases of pulse crops in Uttar Pradesh. A survey of viral diseases of pulse crops in Uttar Pradesh.
- Parmar, H.C., Makani, A.Y., Pali, V., Adsul, H.R., Parmar, D.J. and Patel, K.V., 2022. Assessment of genotypes x environment interaction of black gram (*Vigna mungo* (L.) Hepper) using multivariate analysis. *International Journal of Plant & Soil Science*, **34(23):** 471-477. [Cross Ref]
- Plant Breeding Tools (PBTools). Version: 1.3.) 2013 2020. (c) Copyright International Rice Research Institute (IRRI. http://bbi.irri.org)
- Singh, S. K., Gupta, B. R. and Chib, H. S. 1995. Relation of plant age with yellow mosaic virus infection in urdbean. *Integrated disease management and plant health. Scientific Publishers, Joypur*, 91-92.
- Sridhar, V., Rao, P. J. M., Saikiran, V., Kishore, N. S., Reddy, M. R. and Kumar, G. P. 2023. Development of stable blackgram [*Vigna mungo* (L.) Hepper] genotypes by deciphering genotype× environment interaction using Eberhart-Russell and AMMI models. *Electronic Journal of Plant Breeding*, **14(1)**: 52-59. [Cross Ref]
- USDA National Nutrient Database, 2018. https://ndb.nal. usda.gov/ndb
- Vinothini, R. and Kaleeswari, R. K. 2022. Screening of Blackgram Genotypes for Sulphur Utilization Potential in VerticUstropept. *Madras Agricultural Journal*, **109**(march (1-3)), 1. [Cross Ref]
- Yan, W. 1999. A study on the methodology of yield trial data analysis—with special reference to winter wheat in Ontario. *PhD. diss., University of Guelph, Guelph, Ontario, Canada.*

- Yan, W. 2001. GGE biplot—A Windows application for graphical analysis of multi-environment trial data and other types of two-way data. Agronomy journal, 93(5): 1111-1118. [Cross Ref]
- Yan, W., Hunt, L.A., Sheng, Q. and Szlavnics, Z., 2000. Cultivar evaluation and mega-environment investigation based on the GGE biplot. Crop science, 40(3): 597-605. [Cross Ref]
- Yan, W., Kang, M.S., Ma, B., Woods, S. and Cornelius, P.L. 2007. GGE biplot vs. AMMI analysis of genotypeby-environment data. *Crop science*, **47(2)**: 643-653. [Cross Ref]
- Zhou, C.J., Tian, Z.Y., Li, J.Y., Yang, L., Wu, Y.K., Du, Z.Q., Tang, J.H. and Shi, C., 2011. GGE-Biplot analysis on yield stability and testing-site representativeness of soybean lines in multi-environment trials. *Soybean Sci*, **30(2)**: 318-321.
- Zobel, R.W., Wright, M.J. and Gauch Jr, H.G., 1988. Statistical analysis of a yield trial. *Agronomy Journal*, **80(3)**: 38. [Cross Ref]