Electronic Journal of Plant Breeding



Research Article

Comparison of residual heterosis in F₂ and F₃ segregating generations of Indian Mustard [*Brassica juncea* (L.) Czern & Coss.] crosses

Avralima Sarkar¹, Suvendu Kumar Roy^{1*}, S. Vishnupriya¹, Moumita Chakraborty¹, Lakshmi Hijam¹, Naderla Umamaheswar¹, Sanghamitra Rout², Shivani Bharti³, Bandan Thapa⁴ and Saikat Das⁵

¹Department of Genetics and Plant Breeding, UBKV, Pundibari, Cooch Behar, West Bengal, India

²Department of Genetics and Plant Breeding, CUTM, Paralakhemundi, Odisha, India

³UCO Bank, Suraj Garaha, Lakhi Sarai, Bihar, India

⁴Regional Research Station, Kalimpong, UBKV, West Bengal, India

⁵AICW&BIP, Directorate of Research, UBKV, Pundibari, Cooch Behar, West Bengal, India

*E-Mail: suvendukumarroy@gmail.com

Abstract

The present investigation was undertaken to study the residual heterosis in F_2 and F_3 populations of Indian mustard [*Brassica juncea* (L.) Czern & Coss.]. The study material included eight parents and the 28 crosses of Indian mustard in two generations, namely F_2 and F_3 . The experiment was conducted over two years at Uttar Banga Krishi Viswavidyalaya, Cooch Behar, West Bengal, during *Rabi* season 2020-21 and 2021-22. Out of the 28 crosses, three crosses namely, Npj-194 × DRMR-15-16, Npj-194 × RW-85-59 (Sarma) and Npj-194 × SKJM-05 were found to show significantly positive residual standard heterosis over the standard variety (SKJM-05) for both the characters siliquae per plant and seed yield per plant. This provides us evidence that by improving siliquae per plant, the character seed yield per plant can be improved. The promising cross populations which have shown significant standard heterosis for seed yield in F_2 and F_3 , can be utilized for the development of hybrid varieties, which would be very much beneficial to the marginal farmers of our country as they would be able to use the F_2 and F_3 seeds of the hybrid varieties without any compromise with seed yield.

Keywords: Residual relative heterosis, Residual heterobeltiosis, Residual standard heterosis, Indian Mustard, Segregating generations

INTRODUCTION

Indian mustard [*Brassica juncea* (L.) Czern & Coss.] is the most important oilseed crop, occupying a leading position in the Indian oilseed scenario and playing a crucial role in the country's oilseed economy. It accounts for 20.8% of the world's cultivated area and 10% of global production. India has an ever-increasing population. The standard of living is also rising, resulting in higher per capita consumption of edible oil. It has been estimated that by 2030, the annual growth in demand for edible oil would be 3.54% and accordingly, per capita consumption of edible oil would increase to 23.1 kg/year. Thus, to achieve self-sufficiency in edible oil, 34.1 million tonnes of edible oil would be required, which is equivalent to about 102.3 million tonnes of oilseeds (DRMR 2011). By the end of fiscal year 2022, India produced more than 11 million tons of rapeseed and mustard (Statista, 2022). There is a growing demand for mustard oil in India which is mainly driven by the emerging food industry due to

https://doi.org/10.37992/2024.1503.091

increasing consumer awareness of the numerous health benefits associated with the use of mustard oil, like its anti-bacterial, anti-fungal, anti-viral, anti-inflammatory and most importantly, its anti-carcinogenic properties (Kaur *et al.*, 2019).

Therefore, attainment of high production and productivity of Indian mustard is one of the major objectives in the country. The approach of heterosis breeding has been found to be one of the most successful options that has been employed for the improvement of Brassica varieties for quality and quantity of seed yield and other yield related parameters. With the help of heterosis studies, superior cross combinations of promising hybrids can be identified for developing high yielding varieties (Meena et al., 2015). Residual heterosis occurs when the vield of the segregating generation in some hybrids is likely to be higher than the check variety (Fu et al., 2014). It is the amount of heterosis shown by F2 and its subsequent segregating generations. According to Burton and Brownie (2006), potential selection of hybrid crosses in the F₂ and F₃ generations might be to produce a greater degree of high yielding pure lines than other breeding methods that ignore the possibility of dominance. Hence, identification of better progeny in the early generation is essential for the potential selection process. According to Koseoglu et al. (2017), better progeny than that of the average of the parents at F_{2/3} segregating populations are known as fruitful heterosis (HF). The residual heterosis in F₂ and F₃ populations over the parents is a desirable character and an indication that the superior performance of the F, generation is also reflected in later segregating generations, which indicates that the farmers who once buy seeds of a hybrid mustard variety can keep using the seeds for the next two-three generations without any significant reduction in yield, ultimately, reducing the cost of production of the crop (Duvick et al., 1999). This is applicable only with respect to the hybrid varieties prevalent in the market.

The study of residual heterosis has been exploited in several crops such as chickpea (Adak *et al.*, 2017), wheat (Rajane *et al.*, 2022), okra (Sabesan *et al.*, 2016), Soybean (Bhartiya *et al.*, 2014), rice (Balat *et al.*, 2018), cucumber (Kumari *et al.*, 2021), groundnut (Byadagi *et al.*, 2018), brinjal (Varma *et al.*, 2020) etc. But a very few studies were done on residual heterosis of Indian mustard. Therefore, the present study is an attempt to find the existence of residual heterosis that can be present in F_2 and F_3 populations of Indian mustard so that the seeds of these populations can be used by our farmers without any significant reduction in yield.

MATERIALS AND METHODS

The present study was conducted over two years at Uttar Banga Krishi Viswavidyalaya, Pundibari, Cooch Behar, West Bengal, during *Rabi* seasons 2020-21 and 2021-22. The experimental material for the study included eight parents and the 28 crosses of Indian mustard [*Brassica* juncea (L.) Czern & Coss.] in two generations, namely F₂ and F₂. The crosses , based on inter-varietal hybridization, were obtained from a previous study made by Rout et al. (2021) during the rabi season of 2018-19. The F₂ seeds were sown to develop F₂ populations during Rabi 2020-21. Subsequently, F3 seeds were sown to develop F3 populations during the following year 2021-22. The F₂ and F₂ populations were evaluated along with eight parents for the two consecutive years, in Rabi 2020-21 and 2021-22. Data was recorded from five randomly selected plants from each parent, 20 plants of the 28 F₂ populations and 10 plants from the 28 $\rm F_{_3}$ populations per replicate. Observations were recorded for six morphological characters such as plant height (cm), primary branches per plant, secondary branches per plant, siliquae per plant, 1000 seed weight (g) and seed yield per plant (g). Morphological characters like plant height, primary number of branches, secondary number of branches etc. were found to exhibit significant and positive association with seed yield in a study conducted by Patel et al. (2006). The generation from F₂ to F₃ was advanced by bulking the seed in each of the 28 cross populations separately.

Residual relative heterosis, heterobeltiosis and standard heterosis (using SKJM-05 as the check variety) were estimated as the percentage of deviation of generation mean of F_2 (or F_3) from the mean parental value, better parental value and the standard variety, respectively. For the estimates of residual heterosis (F_2 and F_3) the data were calculated manually. The following formulae suggested by Rao *et al.* (1980) were followed for the calculation of different estimates of heterosis.

Estimation of residual heterosis:

Residual relative heterosis (%) = [{mean of F_2 or F_3 – mean of parent}/ mean of parent] ×100

Residual heterobeltiosis (%) = [{mean of F_2 or F_3 – mean of better parent}/ mean of better parent] ×100

Residual standard heterosis (%) = [{mean of F_2 or F_3 – mean of standard variety}/ mean of standard] ×100

Further, the statistical significance of all the estimates of heterosis was calculated through 't' test formulae suggested by Turner (1953).

RESULTS AND DISCUSSION

Results from ANOVA (**Table 1**) showed that the 36 entries consisting of eight parents and 28 F_2 populations in 2022-21 and F_3 populations in 2021-22 differed significantly for all the six characters in the present study, indicating the presence of large variability among the entries in both the F_2 and F_3 generations. The considerable variability between parents and the segregating population was also noted by Ara *et al.* (2013), in crosses between cultivars of *Brassica* species.

For the character plant height in F2, only six crosses

Sources of	Degrees					Me	an sum	of squares						
variation	of freedom	Plant	height	Primary branches per		branch	Secondary branches per		Siliquae perplant		1000 Seed Weight		Seed yield per plant	
		(ci	m)	pla	ant	pla	ant			(9	g)	(9	g)	
		F_2	F_3	F_2	F ₃	F_2	F_3	F_2	F ₃	F_2	F_3	F_2	F ₃	
Replication	2	1227.715**	339.360	0.456	0.227	21.175**	3.473	3548.591*	* 342.578	1.072	0.135	5.828**	1.550	
Genotype	35	263.789**	1051.315**	0.953**	4.094**	12.727**	7.480**	1272.174*	*2712.316**	1.491**	0.814**	2.346**	4.613**	
Error	70	76.024	245.627	0.241	0.697	2.904	3.191	373.024	846.026	0.382	0.188	0.610	1.845	

Table 1. Analysis of variance for seed yield and its attributing characters in Indian Mustard in F₂ (2020-21) and F₃ (2021-22)generations

** Significant (P \leq 0.01); *Significant (P \leq 0.05), PH = Plant height (cm), PBPP = Primary branches per plant, SBPP= Secondary branches per plant, SPP = Siliquae per plant, 1000 SW = Thousand seed weight (g), SYPP = Seed yield per plant (g)

showed significant residual relative heterosis ranging from -16.17% (Giriraj × RNWR- 09-3) to 18.64% (DRMR-15-16 × Kranti) (Table 2). Significantly positive residual relative heterosis was exhibited by only two crosses, namely, DRMR-15-16 × Kranti and Npj-194 × DRMR-15-16 . In F., 16 crosses showed significant residual relative heterosis that ranged from -33.04 (RNWR-09-3 × PHR-2) to 33.64 (DRMR-15-16 × RNWR-09-3). Out of them, only one cross DRMR-15-16 × RNWR-09-3 (33.64%) expressed significantly positive residual relative heterosis. In F₂, 11 crosses showed significant residual heterobeltiosis that ranged from -20.41% (Giriraj × RNWR-09-3) to 17.65% (DRMR-15- 16 × Kranti). Significantly positive residual heterobeltiosis was shown by only two crosses, namely, Npj-194 × DRMR-15-16 and DRMR-15-16 × Kranti. In F., 22 crosses expressed significant residual heterobeltiosis ranging from -39.87% (SKJM-05×PHR-2) to -22.07% (SKJM-05×Kranti). However, none of the crosses exhibited significantly positive residual heterobeltiosis. In F₂, five crosses showed significant residual standard heterosis ranging from -15.75% (Giriraj × RNWR-09-3) to 15.98% (DRMR-15-16 × Kranti). On comparing all the residual heterosis for plant height, it was revealed that F₂ performed lower than F₂ for residual relative heterosis and heterobeltiosis, whereas for standard heterosis, F₃ performed better than the F, indicating the potential of the segregating population.

For the character primary branches per plant in F_2 , five crosses expressed significant residual relative heterosis ranging from -32.17% (Npj-194 × PHR-2) to 22.98% [RW-85-59 (Sarma) × Giriraj] (**Table 2**). Significantly positive residual relative heterosis over the mid-parental value was exhibited by only one cross RW-85-59 (Sarma) × Giriraj (22.98). Similarly, in F_3 , 18 crosses showed significant residual relative heterosis ranging from 49.6% (SKJM-05 × Kranti) to 194.74% (Npj-194 × DRMR-15-16) and all of them expressed significantly positive residual relative heterosis. In F_2 , 11 crosses exhibited significant residual heterobeltiosis ranging from -75.4% (Npj-194 × Giriraj) to -16.82% [Npj-194 × RW-85-59 (Sarma)]. However, none

of the them were found to express significantly positive residual heterobeltiosis. Similarly, in F₃, 15 crosses had expressed significant residual heterobeltiosis ranging from 48.33% [RW-85-59 (Sarma) × DRMR-15-16] to 172.37% (Npj-194 × DRMR-15-16) and all of them showed significantly positive residual heterobeltiosis. In F₂, the crosses exhibited neither significantly positive residual standard heterosis nor significantly negative residual standard heterosis over the standard variety. In F₂, 20 crosses showed significant residual standard heterosis that ranged from 55.60% (RNWR-09-3 × PHR-2) to 180% [Npj-194 × RW-85-59 (Sarma)] and all of them exhibited significantly positive residual standard heterosis. Out of which Npj-194 × DRMR-15-16 had shown the highest positive residual standard heterosis of 180% over the standard variety SKJM-05. Overall, for all the residual heterosis, F₃ performed better than the F₂.

For the character secondary branches per plant, 17 crosses expressed significant residual relative heterosis in F₂ that ranged from -64.95% (Npj-194 × PHR-2) to 80.23% (Kranti × RNWR- 09-3) (Table 2). Out of which, seven crosses showed significantly positive residual relative heterosis in F₃, 10 crosses expressed significant residual relative heterosis ranging from -51.72% (SKJM-05× Giriraj) to -29.52% [RW-85-59 (Sarma) × Giriraj]. Out of them, none of the crosses were found to express significantly positive residual relative heterosis. In F₂, 16 crosses expressed significant residual heterobeltiosis ranging from -69.02% (Npj-194 × PHR-2) to 75.05% (Kranti × RNWR-09-3). Only two crosses, namely, Kranti × Giriraj and Kranti × RNWR- 09-3 exhibited significantly positive residual heterobeltiosis. In F₃, 12 crosses expressed significant residual heterobeltiosis ranging from - 63.9% (SKJM-05 × Giriraj) to -34.51% [RW-85-59 (Sarma) × Giriraj]. However, none of the crosses were found to exhibit significantly positive residual heterobeltiosis. In F₂, nine crosses exhibited significant residual standard heterosis ranging from 78.9% (RNWR-09-3× PHR-2) to 203.42% (Kranti × PHR-2). All of these crosses expressed significantly positive residual standard Table 2. List of crosses exhibiting positive and negative relative heterosis in F2 and F₃ for the six yield attributing characters in mustard

D											Avralin	na	Sarkar et al.,
	No. of crosses	4	15	6	22	3	0	4	0	11	0	0	0
terosis	No. of Significantly Negative (-) rosses	R-W-85-59 × PHR-2, DRMR-15-16 × Giriraj, Giriraj × RNWR-09-03 and Giriraj × PHR-2	Npj-194× DRMR-15-16, Npj-194× Kranti, Npj-194× RNWR-09-03, Npj-194× PHR-2, RW-85-59(Sarma)× RNWR-09-03, RW-85-59(Sarma)× PHR-2, DRMR-15-16× Kranti, DRMR-15-16× PHR-2, SKJM-05× PHR-2, Kranti× Giriraj, Kranti×RNWR-09-03, Kranti× PHR-2, Giriraj× RNWR-09-03, Giriraj× PHR-2 and RNWR-09-03× PHR-2	Npj-194× PHR-2, RW-85-59(Sarma)× PHR-2, DRMR-15-16×Ciriraj, DRMR- 15-16×PHR-2, SKJM-05× Giriraj, SKJM-05× PHR-2, Giriraj× RNWR-09-03, Giriraj× PHR-2 and RNWR-09-03× PHR-2	Npj-194× RN-85-59, Npj-194× DRMR-15-16, Npj-194× Kranti, Npj-194× RNWR-09-03, RNWR-09-03, RN-85-59× Kranti, RN-85-59× Kranti, RN-85-59× RN-05-03, RN-85-59× PHR-2, DRMR-15-16× SKJM-05× Kranti, SKJM-05× Giriraj, KJJM-05× RNWR-09-03, DRMR-15-16× PHR-2, SKJM-05× Kranti, SKJM-05× Giriraj, KJJM-05× RNWR-09-03, SKJM-05× PHR-2, Kranti× Giriraj, Kranti× RNWR-09-03, SKJM-05× PHR-2, Giriraj, Stanti× PHR-2, Giriraj RNWR-09-03, Giriraj× PHR-2 and RNWR-09-03× PHR-2, DHR-2 and RNWR-09-03× PHR-2, SKJM-05× PHR-2, Giriraj× PHR-2 and RNWR-09-03× PHR-2 AND	Npj-194× RW-85-59(Sarma), RW-85-59(Sarma)× Kranti and Giriraj× RNWR-09-03		Npj-194× Kranti, Npj-194× PHR-2, RW-85-59(Sarma)× PHR-2 and DRMR- 15-16× PHR-2		Npj-194× RW-85-59(Sarma), Npj-194× DRMR-15-16, Npj-194× Kranti, Npj- 194× Giriraj, Npj-194× RNWR-09-03, Npj-194× PHR-2, RW-85-59(Sarma)× PHR-2, DRMR-15-16× PHR-2, SKJM-05× PHR-2, Kranti × PHR-2 and RNWR- 09-03× PHR-2			
Direction of Heterosis	No. of crosses	2	-	2	0	2	0	-	10j- 16, nti,	0	15 1)× 05, anti		19-1- 15- ~ 20 지구- 20
	Significantly Positive (+)	DRMR-15-16 × Kranti (18.64 %) and NPJ-194 × DRMR-15-16(12.06 %)	DRMR-15-16 × RNWR-09-03 (33.64%)	DRMR-15-16× Kranti (17.65%) and Npj-194×DRMR-15-16 (10.02%)		DRMR-15-16× Kranti (15.98%) and Npj-194× Kranti (9.98%)		RW-85-59(Sarma)× Giriraj (22.980%)	Npi-194× DRMR-15-16 (194.74%), Npi-194× RW-85-59(Sarma), Npi-194× SKJM-05, Npi- 194× Giriraj, Npi-194× RNWR-09-03, Npi-194× PHR-2, RW-85-59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05, RW-85-59(Sarma)× Kranti, RW-85-59(Sarma)× Giriraj, DRMR-15-16× SKJM-05, DRMR-15-16× Kranti, DRMR-15-16× Giriraj, SKJM-05× Kranti, SKJM-05× RNWR-09-03, SKJM-05× PHR-2, Kranti× Giriraj and Kranti× PHR-2		Npj-194 × DRMR-15-16 (172.37%), Npj-194× RW-85-59(Sarma), Npj-194× SKJM-05, Npj- 194× Giriraj, Npj-194× PHR-2, RW-85-59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05, RW-85-59(Sarma)×Kranti, RW-85-59(Sarma)× Giriraj, DRMR-15-16× SKJM-05, DRMR-15-16× Kranti, DRMR-15-16× Giriraj, SKJM-05× PHR-2, Kranti × Giriraj and Kranti × PHR-2		Npj-194× DRMR-15-16 (180.00%), Npj-194× RW-85-59(Sarma), Npj-194 × SKJM-05, Npj- 194 × Giriraj, Npj-194 × RNWR-09-03, Npj-194× PHR-2, RW-85-59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05, RW-85-59(Sarma)× Kranti, RW-85-59(Sarma)× Giriraj, RW- 85-59(Sarma)× SKJM-05, RW-85-59(Sarma)× Kranti, RW-85-59(Sarma)× Giriraj, RW- 16× Giriraj, SKJM-05× RNWR-09-03, SKJM-05× PHR-2, Kranti × Giriraj, Kranti × RNWR- 00, 05 × 7-54: D = 20-10 × 50-10 × 50-10 × 50-10 × 10 × 10 × 10 × 10 × 10 × 10 × 10
Generation		ц	٣	s T	щ	т	Ŀ	щ	щ	µ_ م	щ	ц	ίμ ^ω
iype or	residual heterosis	it Residual relative	heterosis	Residual heterobeltiosis		Residual standard	heterosis	Residual errelative	heterosis	Residual heterobeltiosis		Residual	standard heterosis
Character		Plant height (cm)						Primary Residua branches per relative	plant				

Avralima Sarkar et al.,

Avralima Sarkar et al.,

Methods Explored in the interval significanty produce (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Significanty Negatore (+) Methods Significanty Negatore (+) No. of Si	Character Tyne of	Tyne of	Generati	Generation Direction of Heterosis		
F Kranit x FURX-00-03 (30.23%), DRMR-15-16x Kranit, Nei-194x DBMR-15-16, Net-194x Climal, Nei-194x Climal, St.Nuo55 PHR2, Climal ret Nei-1940, St.Nu-055 PHR2, Climal ret		residual heterosis		(+	No. of	Significantly Negative (-) No. of crosses
File 0 NI-1944-Sel3(Sarma): FRUM-55-Sel3(sarma): FRUM-55-	ndary shes	Residual relative heterosis	ц°	Kranti × RNWR-09-03 (80.23%), DRMR-15-16× Kranti, DRMR-15-16× Giriraj, SKJM-05× Kranti, Kranti × Giriraj, Kranti × PHR-2 and Giriraj × PHR-2	~	Npj-194× DRMR-15-16, -194× Giriraj, Npj-194× PH <u>RW-85-59(Sarma)× Kranti</u> PHR-2
Residual F Kranit x RNWR-09-03 (75.05%) and Kranit x Giriaj (66.39%) Z Nupl-194x Rentl, Nupl-194x Giriar Nearly StrAm-51-16. Reschellesis Np1-194x Mirez, RWA5-56(Sarma)x DRMR-15-16. Np1-194x Rentl, Np1-194x Giriar P. RP2, DRMR-15-16. F Np1-194x Rinez, RWA-05-03 (75.05%) and Kranit x Giriaj (66.39%) Z Np1-194x Giria, RWA5-56(Sarma)x DRMR-15-16. F Np1-194x Giria, RWA5-56(Sarma)x DRMR-15-16. S(Sarma)x StMM-05- PHR2, Ciriar), RWA5-56(Sarma)x Kranit, 56(Sarma)x StMM-05- PHR2, 2000-03. F Non-1 S(KM-05- NHR2, Ciriar), S(KM-05- NHR2, RWA5-56)(Sarma)x Kranit, 56(Sarma)x StMM-05- NHR2, 2000-03. Residual F Kranit x PHR2, 2(03.42%), RW.45-59(Sarma)x Giriar), S(MM-05- NHR2, 2000-03. Residual F Kranit x RWK-09.03 and RWW-09-03. Residual F Kranit x RWK-09.03 and RWW-09-03. Residual F NWR-05- NHR2, Kranit x RWW-09-03 and Giriar) x PHR2, 2. Residual F NWR-05- NHR2, Kranit x RWW-09-03 and RWW-09-03. Residual F Kranit x RWW-09-03 and RWW-09-03. Residual F Kranit x RWW-09-03. Residual F Kranit x RWW-09-03. Residual F Kranit x RWW-09-03. <td></td> <td></td> <td>ц"</td> <td></td> <td>0</td> <td>Npj-194× Giriraj, RW-85-59(Sarma)× SKJM-05, RW-85-59(Sarma)×10 Kranti, RW-85-59(Sarma)× Giriraj, RW-85-59(Sarma)× RNWR-09-03, RW-85-59(Sarma)× PHR-2, SKJM-05× Giriraj, Kranti × RNWR-09- 03, Giriraj × RNWR-09-03 and Giriraj × PHR-2</td>			ц"		0	Npj-194× Giriraj, RW-85-59(Sarma)× SKJM-05, RW-85-59(Sarma)×10 Kranti, RW-85-59(Sarma)× Giriraj, RW-85-59(Sarma)× RNWR-09-03, RW-85-59(Sarma)× PHR-2, SKJM-05× Giriraj, Kranti × RNWR-09- 03, Giriraj × RNWR-09-03 and Giriraj × PHR-2
Fa 0 00):194 clirinaj. RNR-15-16, 16 Selamaja PIRVa SKJM-05 Selamaja Cirinaj. RNR-15-16, 16 Selamaja PIRVa SKJM-05 Selamaja Cirinaj. RNM-15-16 Selamaja PIRVa SKJM-05 Selamaja Cirinaj. RNM-15-16 Selamaja PIRVa SKJM-05 Selamaja Cirinaj. RNM-15-16 Selamaja PIRVa SKJM-05 Selamaja Cirinaj. SKJM-05 Selamaja PIRVa SKJM-05 SKJM-05 Selamaja Cirinaj. Selamaja PIRVa SKJM-05 SKJM-05 SKJM-05 Selamaja PIRVa SKJM-05 SKJM-05 SKJM-05 Selamaja PIRVA SKJM-05 SKJM-05 SKJM-05		Residual heterobeltiosi	ц ² 2	Kranti × RNWR-09-03 (75.05%) and Kranti × Giriraj (66.38%)		Npj-194× RW-85-59(Sarma), Npj-194× DRMR-15-16, Npj-194×14 SKJM-05, Npj-194× Kranti, Npj-194× Giriraj, Npj-194× RNWR-09- 03, Npj-194× PHR-2, RW-85-59(Sarma)× DRMR-15-16, RW-85- 59(Sarma)× SKJM-05, RW-85-59(Sarma)× PHR-2, DRMR-15-16× PHR-2, SKJM-05× PHR-2, Giriraj × PHR-2 and RNWR-09-03× PHR- 2
Residual F. Kranit SHR.2 (30 42%), RW.65-59(Sarma)× Giriraj, 9 standard F SKUM-05× Kranit, DRMR-15-16× Kranit, DRMR-15-16× Kranit, SKUM-05× SARM-05 Npi-194× SKUM-05× Kranit, SKUM-05× SRUM-05 Npi-194× SKUM-05× Kranit, SKUM-05× SRUM-05 SKUM-05× Kranit, SKUM-05× RAVK-05-003, Kranit SSKUM-05× Kranit, SKUM-05× RAVK-05-03, Kranit SSKUM-05× Kranit, SKUM-05× RAVK-09-03, Kranit SSKUM-05× SRUM-05× RAVK-09-03 SSKUM-05× RAVK-09-03× CRAVK-09× SSG(Sarma)× SKUM-05, Npi-194× Kranit, × Giriraj, Npi-194× SKUM-05 SKUM-05× CRAVK-09-03× CRAVK-09-03 SSKUM-05× SKUM-05× SKUM-05× SKUM-05× SKUM-05× Npi-194× SKUM-05× Npi-194× SKUM-05× Npi-194× SKUM-05× SKUM-			щ		0	Giriraj, RW-85-59(Sarma)× DRMR-15-16, SKJM-05, RW-85-59(Sarma)× Kranti, Giriraj, RW-85-59(Sarma)× RNWR-09-03, PHR-2, 6× Giriraj, SKJM-05× Giriraj, Kranti × RNWF VR-09-03 and Giriraj × PHR-2
Fs SKUM-05× Kranti, SKUM-05× PHR-2, Kranti X Girrigi, Kranti X Girrigi, Kranti X Girrigi, Kranti X KumCu9-03 and RWR-09-03× PHR-2, Kranti X Girrigi (64.28%) 3 Residual Fz Nipi-194× SKUM-05 (66.28%), Kranti X Girrigi (64.28%) 7 Nipi-194× SKUM-05 Nipi-194× SKUM	_ •	Residual standard	ц г	Kranti × PHR-2 (203.42%), RW-85-59(Sarma)× Giriraj, DRMR-15-16× Kranti, DRMR-15-16× PHR-2,		
Nbj-194× DRMR-15-16 (64.28%), Kranti × Giriraj (64.28%) ae per Residual F Kanti St.JM-05 (56.25%) DRMR-15-16 × St.JM-05, St.JM - 5 St.JM-05 and RW-85- and Npj-194× SLM-05 (56.25%) ae per Residual F Kranti Stringi (59.31%), DRMR-15-16 × St.JM-05, Kranti Sol Stranti St.JM-05 and RW-85- and Npj-194× SLM-05 × RLM-05, St.JM - 5 Npj-194× SLM-05 × RLM-05 St.JM-05 and RW-85- St.JM-05 × RLM-05 Npj-194× SLM-05 St.JM-05 and RW-85- St.JM-05 × RLM-05 St.JM-05 and RW-85- St.JM-05 × RLM-05 St.JM-05	_	heterosis	டீ	SKJM-05× Kranti, SKJM-05× PHR-2, Kranti × Giriraj, Kranti × RNWR-09-03 and RNWR-09-03× PHR-2		
ae per Residual F ₂ Krantis Giriraj (59.31%), DRMR-15-16× SKJM-05, SKJM-05, Npi-194× SKJM-05, Npi-194× PHR-2, RW-85- relative 05× Kranti, SKJM-05 × NHR-2 × PHR-2 and RNWR-09-03× Rranti × RNWR-09-03, Kranti 59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05 and RW- heterosis × Npi-194× SKJM-05 (84.24%), Npj-194× RW-85-59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05 and RW- heterosis × Npi-194× SKJM-05 (84.24%), Npj-194× RW-85-59(Sarma), S Npj-194× DRMR-15-16, SKJM-05 × RW-85-59(Sarma), S Npj-194× DRMR-15-16, SKJM-05 × RW-85-59(Sarma), S Npj-194× DRMR-15-16, SKJM-05 × RW-85-59(Sarma)× DRMR- heterobelticosis 15-16, DRMR-15-16, SKJM-05 × Kranti, Residual F ₂ SKJM-05 × SKJM-05 × Kranti, Residual F ₂ SKJM-05 × SKJM-05 × Kranti, Npi-194× SKJM-05 × SKJM-05 × Kranti, Residual F ₂ SKJM-05 × SKJM-05 × Kranti, Npi-194× SKJM-05 × SKJM-05 × Kranti, Residual F ₂ SKJM-05 × SKJM-05 × Kranti, Residual F ₂ SKJM-05 × SKJM-05 × Kranti, Residual F ₂ SKJM-05 × RM-05 × Kranti, Residual F ₂ SKJM-05 × RM-05 × SKJM-05 × SKJM-05, SKJM Npj-194× SKJM-05 × PHR-2 SKJM-05 × RMW-09-03 × DHR-2 Npi-194× SKJM-05 × RMW-09-03, Kranti × RW-85-59(Sarma)× PHR-2 Npi-194× SKJM-05 × SKJM-05 × SKJM-05, SKJM Npi-194× SKJM-05 × RMW-09-03 × RTR-15-16, SKJM-05, SKJM Npi-104× 05 × SKJM-05 × SKJM-05, SKJM Npi-104× SKJM-05 × RMW-09-03 × DHR-2 Npi-104× SKJM-05 × RMW-09-03 × DHR-2 Npi-104× SKJM-05 × RMW-09-03 × RMR-15-16, SKJM-05, SXJM Npi-104× 05 × RMW-15-16, SKJM-05, SXJM Npi-104× 05 × RMW-09-03 × DHR-2				64.28%), Kranti × Giriraj (64 (56.25%)		0
Fa Npj-194× SKJM-05 (84.24%), Npj-194× RW-85-59(Sarma), 5 5 Npj-194× SKJM-05 (84.24%), Npj-194× RW-85-59(Sarma), 5 Npj-194× SKJM-05 (84.24%), Npj-194× RW-85-59(Sarma), 5 RND: 194× DRMR-15-16, SKJM-05× RNWR-09-03 NDj-194× SKJM-05, NDj-194× Kranti, × Giriraj, NDj-194× PHR-2 F SKJM-05× PHR-2 (200.00%), RW-85-59(Sarma)× DRMR- 8 NDj-194× SKJM-05 F SKJM-05× PHR-2 (200.00%), RW-85-59(Sarma)× DRMR- 8 NDj-194× SKJM-05 F Npj-194× SKJM-05 SKJM-05× SKJM-05 F Npj-194× SKJM-05 T RNWR-09-03× PHR-2 RW-85-59(Sarma)× SKJM-05 PHR-2 RNWR-09-03× PHR-2 RW-85-59(Sarma)× SKJM-05 SKJM-05× SKJM-05 F Npj-194× SKJM-05 T RW-85-59(Sarma)× PHR-2 F Npj-194× SKJM-05× RNWR-09-03 Kranti × RNWR-09-03 SKJM-05× SLM-05 F Npj-194× SKJM-05× RNWR-09-03 Kranti × Chiraj (60.60%), DRMR-15-16× SKJM-05 SKJM-05× SLM-05 F Kranti × Giriraj (60.60%), DRMR-15-16× SKJM-05 Npj-194× SKJM-05 SKJM-05× SLM-05 F Kranti × Giriraj (60.60%), DRMR-15-16× SKJM-05 Npj-194× SKJM-05× SLM-05 SKJM-05× SLM-05 F Kranti × Giriraj (ae per	Residual relative heterosis	Ъ2	Krantix Giriraj (59.31%), DRMR-15-16× SKJM-05, SKJM- 05× Kranti, SKJM-05× PHR-2, Kranti × RNWR-09-03, Kranti × PHR-2 and RNWR-09-03× PHR-2	~	Npj-194× SKJM-05, Npj-194× Giriraj, Npj-194× PHR-2, RW-85-6 59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05 and RW- 85-59(Sarma)× PHR-2
F SkJM-05× PHR-2 (200.00%), RW-85-59(Sarma)× DRMR- 8 Npj-194× SkJM-05, Npj-194× Kranti, × Giriraj, Npj-194× Kranti, × Giriraj, Npj-194× PHR-2 15-16, DRMR-15-16× SkJM-05, SkJM-05, SkJM-05, Kranti, RW-85-59(Sarma)× SKJM-05, and RW-85-59(Sarma)× PHR-2 Kranti × Giriraj, Kranti × RNWR-09-03, Kranti × PHR-2 and RW-85-59(Sarma)× SKJM-05 and RW-85-59(Sarma)× PHR-2 F Nupj-194× SKJM-05, (66.78%), Npj-194× RW-85-59(Sarma), 4 RW-85-59(Sarma)× PHR-2 F Nupj-194× SKJM-05 (76.78%), Npj-194× RW-85-59(Sarma), 4 RW-85-59(Sarma)× PHR-2 F Npj-194× SKJM-05 (76.78%), Npj-194× RW-85-59(Sarma), 4 RW-85-59(Sarma)× PHR-2 F Npj-194× SKJM-05 (76.78%), Npj-194× RW-85-59(Sarma), 4 RW-85-59(Sarma)× PHR-2 F Npj-194× SKJM-05 (76.78%), Npj-194× RW-09-03 Aranti F Npj-194× SKJM-05 (76.78%), Npj-194× RW-09-03 Aranti F Npj-194× SKJM-05 SKJM-05 Aranti F Npj-194× SKJM-05 SKJM-05 SKJM-05 F Npj-194× SKJM-05 SKJM-05 SKJM-05× RNWR-09-03 and RNWR-09-03× PHR-2 Aranti			щ	Npj-194× SKJM-05 (84.24%), Npj-194× RW-85-59(Sarma), Npj-194× DRMR-15-16, SKJM-05× RNWR-09-03 and RNWR-09-03× PHR-2		0
F3 Npj-194× SKJM-05 (76.78%), Npj-194× RW-85-59(Sarma), 4 RW-85-59(Sarma)× PHR-2 Npj-194× DRMR-15-16 and SKJM-05× RNWR-09-03	—	Residual heterobeltiosi	щ Г S	SKJM-05× PHR-2 (200.00%), RW-85-59(Sarma)× DRMR- 15-16, DRMR-15-16× SKJM-05, SKJM-05× Kranti, Kranti × Giriraj, Kranti × RNWR-09-03, Kranti × PHR-2 and RNWR-09-03× PHR-2		Npj-194× SKJM-05, Npj-194× Kranti, × Giriraj, Npj-194× PHR-2,6 RW-85-59(Sarma)× SKJM-05 and RW-85-59(Sarma)× PHR-2
F2 Kranti × Giriraj (60.60%), DRMR-15-16× SKJM-05, SKJM- 7 7 05× Kranti, SKJM-05× PHR-2, Kranti × RNWR-09-03, Kranti x PHR-2 and RNWR-09-03× PHR-2 x PHR-2 × PHR-2 and RNWR-09-03× PHR-2 126.00%), Npj- 6 194×DRMR-15-16, Npj-194× SKJM-05, 2× DRMR-15-16, SKJM-05× RNWR-09-03× PHR-2			Ĕ	Npj-194× SKJM-05 (76.78%), Npj-194× RW-85-59(Sarma), Npj-194× DRMR-15-16 and SKJM-05× RNWR-09-03		RW-85-59(Sarma)× PHR-2
Npj-194×RW-85-59(Sarma) (126.00%), Npj- 6 194×DRMR-15-16, Npj-194× SKJM-05, 2× DRMR-15-16, SKJM-05× RNWR-09-03 and RNWR-09-03× PHR-2		Residual standard heterosis	т ²	Kranti × Giriraj (60.60%), DRMR-15-16× SKJM-05, SKJM- 05× Kranti, SKJM-05× PHR-2, Kranti × RNWR-09-03, Kranti × PHR-2 and RNWR-09-03× PHR-2	2	0
			щ	(Sarma) (126.00%), 94× SKJM-05, 2× DRMR-15-16 and RNWR-09-03× PHR-2		0

Character	Type of residual Gene	Type of residual Generation Direction of Heterosis	
	heterosis	Significantly Positive (+) No. of	Significantly Negative (-) No. of
1000 Seed Weight	I Residual relative F ₂ heterosis	RW-85-59(Sarma)× PHR-2 (84.90%), RW-85-3 59(Sarma)× RNWR-09-03 and DRMR-15-16× Kranti	Npj-194× SKJM-05, Npj-194× PHR-2, RW-85-59(Sarma)× SKJM-05,10 DRMR-15-16× SKJM-05, DRMR-15-16× Giriraj, DRMR-15-16× RNWR- 00 02 eV IM 05× Vravit
(B)	۳	SKJM-05× Giriraj (41.20%), Npj-194× DRMR-15-16,6 Npj-194× SKJM-05, Npj-194× Giriraj, DRMR-15-16×	05-05, Shum-05 Manu, SKJM-05s RNWR-09-03, SKJM-05s PHR-2 and Giriraj × PHR-2 RW-85-59(Sarma)× Kranti
	Residual F ₂ heterobeltiosis		Npj-194× SKJM-05, Npj-194× PHR-2, RW-85-59(Sarma)× SKJM-05,12 RW-85-59(Sarma)× RNWR-09-03, DRMR-15-16× SKJM-05, DRMR-15- 16× Giriraj, DRMR-15-16× RNWR-09-03, SKJM-05× Kranti, SKJM-05× Giriraj, SKJM-05× RNWR-09-03, SKJM- 05× PHR-2 and Giriraj, SKJM-05× Giriraj, SKJM-05× RNWR-09-03, SKJM-
	L.E.	Npj-194× SKJM-05 (48.35%) and SKJM-05× Giriraj2 (33.96%)	RW-85-59(Sarma)× Kranti
	Residual F ₂ standard heterosis	RW-85-59(Sarma)× PHR-2 (48.86%) and Giriraj ×2 RNWR-09-03 (40.14%)	Npj-194× RW-85-59(Sarma), Npj-194× DRMR-15-16, Npj-194× SKJM-23 05, Npj-194× Kranti, Npj-194× RNWR-09-03, Npj-194× PHR-2, RW- 85-59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05, RW-85- 59(Sarma)× Kranti, RW-85-59(Sarma)× Giriraj, RW-85-59(Sarma)× RNWR-09-03, DRMR-15-16× SKJM-05, DRMR-15-16× Giriraj, DRMR- 15-16× RNWR-09-03, DRMR-15-16× PHR-2, SKJM-05× Kranti, SKJM- 05× Giriraj, SKJM-05× RNWR-09-03, SKJM-05× PHR-2, Kranti × RNWR- 09-03, Kranti × PHR-2, Giriraj × PHR-2, Giriraj × RNWR-
	ц Ш	SKJM-05× Giriraj (49.25%), Npj-194× SKJM-05 and3 Npi-194× Girirai	RW-85-59 (Sarma)× Kranti 1
ieed yield p plant (g/plant)	Seed yield per Residual relative F ₂ heterosis plant (g/plant)	Kranti × RNWR-09-03 (51.50%) and SKJM-05×1 Giriraj (50.89%)	Npj-194× RW-85-59(Sarma), Npj-194× SKJM-05, Npj-194× Kranti, Npj-15 194× Giriraj, Npj-194× PHR-2, RW-85-59(Sarma)× SKJM-05, RW-85- 59(Sarma)× Giriraj, RW-85-59(Sarma)× PHR-2, DRMR-15-16× SKJM-05, DRMR-15-16× Kranti, DRMR-15-16× Giriraj, DRMR-15-16× RNWR-09- 03, SKJM-05× RNWR-09-03, Giriraj × RNWR-09-03, Giriraj × PHR-2
	ц. С	Npj-194× DRMR-15-16 (76.86%), Npj-194× RW-85-3 59(Sarma) and DRMR-15-16× Girirai	Girraj × PHR-2 1
	Residual F ₂ heterobeltiosis	Kranti × RNWR-09-03 (43.50%) 1	Npj-194× RW-85-59(Sarma), Npj-194× SKJM-05, Npj-194× Kranti, Npj-20 194× Giriraj, Npj-194× RNWR-09-03, Npj-194× PHR-2, RW-85-59(Sarma)× DRMR-15-16, RW-85-59(Sarma)× SKJM-05, RW-85-59(Sarma)× Kranti, RW-85-59(Sarma)× Giriraj, RW-85-59(Sarma)× RNWR-09-03, RW-85- 59(Sarma)× PHR-2, DRMR-15-16× SKJM-05, DRMR-15-16× Giriraj, DRMR-15-16× RNWR-09-03, SKJM-05× Giriraj, SKJM-05× RNWR-09-03, Kranti × Girirai, Girirai × RNWR-09-03 and Giriraj × PHR-2
	Ľ	0	RW-85-59(Sarma)× Kranti, RW-85-59(Sarma)× Giriraj, RW-85-6 59(Sarma)× RNWR-09-03, DRMR-15-16× PHR-2, SKJM-05× RNWR-09- 03 and SKJM-05× PHR-2
	Residual F ₂ standard heterosis	DRMR-15-16× RNWR-09-03 (46.11%) and Npj-194×2 DRMR-15-16 (41.12%)	Npj-194× SKJM-05, Npj-194× Kranti, Npj-194× PHR-2, RW-85-9 59(Sarma)× PHR-2, DRMR-15-16× SKJM-05, DRMR-15-16× Giriraj, SKJM-05× Girirai. SKJM-05× RNWR-09-03 and Girirai × RNWR-09-03
	Ŀ	DRMR-15-16× Giriraj (45.77%)	

Avralima Sarkar et al.,

https://doi.org/10.37992/2024.1503.091

heterosis. In F_3 , three crosses showed significant residual standard heterosis ranging from 56.25% (Npj-194× SKJM-05) to 64.28% (Npj-194 × DRMR-15-16 and Kranti × Giriraj). All of them exhibited significantly positive residual standard heterosis. Overall, negative residual mid and better parent heterosis were revealed for both the generations and positive residual standard heterosis was observed for F_2 and F_3 generations. In all the cases, F_2 performed superior to the F_3 for the respective character.

For siliquae per plant, in F₂, 13 crosses showed significant residual relative heterosis ranging from -22.19% [RW-85-59 (Sarma) × SKJM-05] to 59.31% (Kranti × Giriraj) (Table 2). Out of which, seven crosses had expressed significantly positive residual relative heterosis. In F₂, five crosses expressed significant residual relative heterosis ranging from 41.05% (RNWR-09-3× PHR-2) to 84.24% (Npj-194× SKJM-05). All of them expressed significantly positive residual relative heterosis with Npj-194 × SKJM-05 as the best performer. In F2, 14 crosses expressed significant residual heterobeltiosis ranging from -22.78% [RW-85-59 (Sarma) × SKJM-05] to 200% (SKJM-05 × PHR-2). Eight of them had expressed significantly positive residual heterobeltiosis with SKJM-05 × PHR-2 as the best performer. In F₃, five crosses showed significant residual heterobeltiosis ranging from -33.79% [RW-85-59 (Sarma) × PHR-2] to 76.78% (Npj-194 × SKJM-05). Out of them, four crosses expressed significantly positive residual heterobeltiosis and only one cross, RW-85-59 (Sarma)× PHR-2 showed significantly negative residual heterobeltiosis. In F2, seven crosses exhibited significant residual standard heterosis ranging from 25.20% (DRMR-15-16 × SKJM-05) to 60.60% (Kranti × Giriraj) and all of them expressed significantly positive residual standard heterosis. In F₂, six of the crosses showed significant residual standard heterosis ranging from 53.95% (RNWR-09-3× PHR-2) to 126.00% [(Npj-194 × RW-85-59 (Sarma)]. All of them showed significantly positive residual standard heterosis. Here, F₃ performed well for all the forms of residual heterosis especially the crosses involving Npj-194 as a parent.

For the character 1000 seed weight, in F₂, 13 crosses expressed significant residual relative heterosis ranging from -36.38% (Npj-194 × SKJM-05) to 84.90% [RW-85-59 (Sarma) × PHR-2] (Table 2) with only three crosses exhibiting significantly positive residual relative heterosis. In F₃, six crosses exhibited significant residual relative heterosis ranging from -27.82% (2×5) to 50.15% (Npj-194 × SKJM-05). Five of them showed significantly positive residual relative heterosis. The other five crosses showed significantly negative residual relative heterosis in F3. In F2, 12 crosses expressed significant residual heterobeltiosis that ranged from -48.00% [SKJM-05, RW-85-59 (Sarma)× RNWR-09-3] to -24.96% (SKJM-05 × Giriraj) and none of them showed significantly positive residual heterobeltiosis. In F₃, three crosses showed significant residual heterobeltiosis ranging from -29.91% Avralima Sarkar et al.,

[RW-85-59 (Sarma) × Kranti] to 48.35% (Npj-194 × SKJM-05). In F_2 , majority of the crosses, i.e., 25 out of 28 crosses exhibited significant residual standard heterosis that ranged from -54.62% [RW-85-59 (Sarma) × RNWR-09-3] to 48.86% [RW-85-59 (Sarma) × PHR-2]. Out of them, only two crosses, namely RW-85-59 (Sarma) × PHR-2 and Giriraj × RNWR-09-3 exhibited significantly positive residual standard heterosis. In F_3 , only four crosses exhibited significant residual standard heterosis ranging from -24.02% [RW-85-59(Sarma) × Kranti] to 49.25% (SKJM-05 × Giriraj). Out of them, three crosses, exhibited significantly positive residual standard heterosis over the standard variety SKJM-05. Overall, F_3 performed better than the F_2 for all the types of residual heterosis.

For the character seed yield per plant in F2, 17 crosses showed significant residual relative heterosis ranging from -61.26% (Npj-194 × SKJM-05) to 51.50% (Kranti × RNWR-09-3) (Table 2). Out of them, only two crosses, namely, SKJM-05 × Giriraj and Kranti × RNWR-09-3 showed significantly positive residual relative heterosis In F₃, only four crosses exhibited significant residual relative heterosis ranging from -31.62% (Giriraj × PHR-2) to 76.86% (Npj-194 × DRMR-15-16). Out of them, three crosses exhibited significantly positive residual relative heterosis. In F2, majority of the crosses, i.e., 21 out of the 28 crosses expressed significant residual heterobeltiosis ranging from -68.67% (Giriraj × RNWR-09-3) to 43.50% (Kranti × RNWR-09-3). However, only one cross Kranti × RNWR-09-3 (43.50%) exhibited significantly positive residual heterobeltiosis over the better parent RNWR-09-3. In F₃, six crosses showed significant residual heterobeltiosis ranging from - 50.46% (DRMR-15-16× PHR-2) to -9.47% (SKJM-05× RNWR-09-3) and none of them showed significantly positive residual heterobeltiosis. In F_2 , 12 crosses exhibited significant residual standard heterosis ranging from -54.52% (Giriraj × RNWR-09-3) to 46.11% (DRMR-15-16 × RNWR-09-3) In F₃, only one cross DRMR-15-16 × Giriraj with significantly positive residual standard heterosis of 45.77% was found. Overall, F3 performed better than the F₂ for residual mid and better parent heterosis whereas, for standard heterosis both the generations performed equally, irrespective of the crosses.

In the present study, the residual relative heterosis, heterobeltiosis and standard heterosis in desired direction were found for all the characters for both the generations. This retention of heterosis may be due to the presence of transgressive segregants and tight linkage between some of the favorable genes that are controlling the characters (Sabesan *et al.*, 2016). There are many causes for residual heterosis. According Kumar *et al.* (2002), low inbreeding depression might be the reason for residual heterosis. It is also found that heterosis was expressed in both the positive and negative directions for most of the characters. The positive and negative expressions indicated the role of dominant and recessive genes in the

inheritance of the characters (Rajane *et al.*, 2022). The **Table 2** shows the crosses that exhibited a positive and negative residual heterosis for the characters for both the generations. Sabesan *et al.* (2016) suggested that negative heterosis in F_2 population indicates that those characters suffer from severe inbreeding depression.

From **Table 2**, it is also evident that some notable crosses in both the generations performed well for given one or more characters such as DRMR-15-16 × Kranti, Npj-194 × DRMR-15-16, Npj-194 × RW-85-59 and Npj-194 × SKJM-05, DRMR-15-16 × Giriraj and Kranti × RNWR-09-3. The number of mustard crosses with positive residual heterosis (relative, heterobeltiosis and standard heterosis) in F_2 and F_3 generations are depicted in **Fig 1**. Hence, out of all the 28 crosses, some of the crosses were chosen based on their superior performances for the characters under study in F_2 and F_3 generations as shown in **table 3**.

Out of the 28 crosses, three crosses namely, Npj-194 × DRMR-15-16, Npj-194 × RW-85-59 (Sarma) and Npj-194 × SKJM-05 were found to show significantly positive residual standard heterosis over the standard variety for two characters namely, siliquae per plant and seed yield per plant. This provides us evidence that by improving siliquae per plant, the character seed yield per plant can be improved. Therefore, it may be inferred that the crosses identified are useful to study the residual heterosis and also to improve the particular character in the breeding programme under present study. Since the development of hybrid seed of Indian mustard is not adequately developed in India, search for the crosses manifesting heterosis largely due to additive gene effects should be attempted to make proper use of residual heterosis.

The present study was done to find out whether residual heterosis is available in Indian mustard and if it can be utilized in the segregating generations. For economic application, similar kind of research can be conducted by including the commercially available hybrid varieties of Indian mustard. The promising cross populations in the present study, which have shown significant standard heterosis for seed yield in F_2 and F_3 , can be utilized for the development of hybrid varieties, which would be very much beneficial to the marginal farmers of our country as they would be able to directly use the F_1 followed by indirectly using the F_2 and F_3 seeds of the promising hybrid varieties, without any significant compromise with seed yield.

ACKNOWLEDGEMENT

The authors are thankful to the Dean, Faculty of Agriculture and Director of Research, Uttar Banga Krishi Viswavidyalaya, Pundibari, Cooch Behar, West Bengal for providing all the infrastructural facilities for conducting the research work.

S. No.	Crosses	Positive residual heterosis in respective generations	Characters
		F ₂	Plant height
1.	DRMR-15-16 × Kranti	F ₃	Primary branches per plant
		F ₂	Secondary branches per plant
		F_2	Plant height
		F ₃	Primary branches per plant
0		F ₃	Secondary branches per plant
2.	Npj-194 × DRMR-15-16	F ₃	Siliquae per plant
		F ₃	1000 seed weight
		F ₃	Seed yield per plant
		F ₃	Primary branches per plant
3.	Npj-194 × RW-85-59 (Sarma)	F ₃	Siliquae per plant
		F ₃	Seed yield per plant
		F ₃	Primary branches per plant
4.	Npj-194 × SKJM-05	F ₃	Siliquae per plant
		F ₃	Seed yield per plant
-		F ₃	Seed yield per plant
5.	DRMR-15-16 × Giriraj	F ₃	Primary branches per plant
		F_2	Secondary branches per plant
6.	Kranti × RNWR-09-3	F_2	Seed yield per plant
		F_2	Primary branches per plant

Table 3. Promising crosses and their residual heterosis performance for the different characters

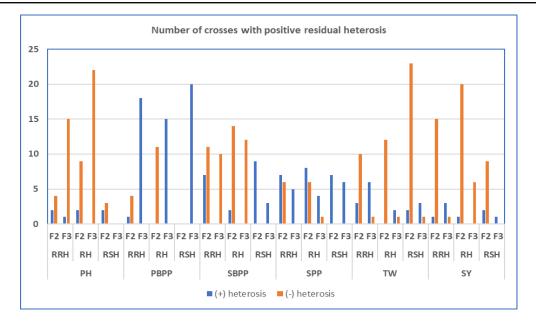


Fig. 1. Number of crosses with positive residual heterosis (relative heterosis, heterobeltiosis and standard heterosis) in F₂ and F₃ generations of mustard

REFERENCES

- Adak, A., Sari, D., Dari, H. and Toker, C. 2017. Gene effects of *Cicer resticulatum* on qualitative and quantitative characters in the cultivated chickpea. *Plant Breed.*, **136**(6): 939-947. [Cross Ref]
- Ara, S., Afroz, S., Noman, M.S., Bhuiyan, M.S.R. and Zia, M.I.K. 2013. Variability, correlation and path analysis in f₂ progenies of inter-varietal crosses of brassica rapa. *J. Environ. Sci. & Natural Resources.*, 6(1): 217 – 220. [Cross Ref]
- Balat, J.R., Thakor, R.P., Delvadiya, I.R. and Rathva, S.R. 2018. Heterosis and inbreeding depression in F_2 population of rice (*Oryza sativa* L.) for yield and related characters. *J. p* h a r m a c o g n . *phytochem.*, **7**(3): 3224-3226.
- Bhartiya, A., Singh, K., Aditya, J.P., Puspendra. and Gupta, M. 2014. Residual relative heterosis and heterobeltiosis for different agro-morphological characters in early segregating generations of soybean [*Glycine max* (L.) Merrill] crosses. *Soybean res.*, **12**(1): 28-35.
- Burton, J.W. and Brownie, C. 2006. Heterosis and inbreeding depression in two soybean single crosses. *Crop Sci.*, **46**:2643-8. [Cross Ref]
- Byadagi, U.R., Venkataravana, P. and Priyadarshini S.K. 2018. Genetic variability study in F₂ and F₃ populations of three crosses of groundnut (*Arachis Hypogaea* L.). *J. pharmacogn. phytochem.*, 7(5): 3139-3143.

- DRMR. 2011. VISION 2030. Directorate of Rapeseed-Mustard Research, Bharatpur, Rajasthan.
- Duvick, D. N. 1999. Heterosis: feeding people and protecting natural resources. Genetics and exploitation of heterosis in crops (eds J.G. Coors and S. Pandey). https://doi.org/10.2134/1999. geneticsandexploitation.c3. [Cross Ref]
- Fu, D., Xiao, M., Alice, H., Fu, Y., Liu, G., Jiang, G. and Zhang, H. 2014. Utilization of crop heterosis: a review. *Euphytica.*,**197**:167-173. [Cross Ref]
- Kaur, R., Sharma, A.K., Rani, R., Mawlong, I. and Rai, P.K. 2019. Medicinal qualities of mustard oil and its role in human health against chronic diseases: A Review. Asian J Dairy Food Res., 38(2):98-104. [Cross Ref]
- Koseoglu, K., Adak, A., Sari, D., Sari, H., Ceylan, F.O. and Toker, C. 2017. Transgressive segregations for yield criteria in reciprocal interspecific crosses between *Cicer* arietinum *L*. and *C. reticulatum* Ladiz. *Euphytica.*, **213**: 1–11. [Cross Ref]
- Kumar, P., Yadav, T.P., Yadav, A.K. and Thakral, S.K. 2002. Estimates of heterosis with respect to yield and its component characters. J. Oil Seeds Res., 5: 72-76.
- Kumari, R., Kumar, R. and Mehta, D.K. 2021. Expression of heterosis and residual heterosis For characters for earliness and yield in cross combinations of cucumber (*Cucumis sativus* L.) developed by introgression of indigenous and exotic sources. *Sci. Hortic.*, **277**: 109781. [Cross Ref]

- Meena, J., Harsha., Pant, U. and Bahjan, R. 2015. Heterosis analysis for yield attributed traits in Indian mustard [*Brassica juncea* (L.) Czern & Coss.]. *Electronic Journal of Plant Breeding*, 6(4): 1103-1107.
- Patel, J.M., Patel, K.M., Prajapati, K.P. and Patel, C.J. 2006. Studies on relationship between yield and its components in Indian mustard. *Madras Agric. J.*, **93**(1-6): 111-115.
- Rajane, A.R., Potdukhe, N.R., Sai Prasad S.V., Gahukar, S.J., Gite, B.D. and Walke, S.R.D. 2 0 2 2 . Assessment of residual useful heterosis for grain yield and yield component characters in the bread wheat (*Triticum aestivum* L.). *J. Pharm. Innov.*, **11**(7): 2926-2930.
- Rao, N. 1980. Statistics for agricultural science. Oxford and IBH publishing company, New Delhi (India).
- Rout, S. 2021. Studies on combining ability for seed yield along with its attributing characters and aphid resistance in mustard [*Brassica juncea* (L.) Czern & Coss.] Department of Genetics and Plant Breeding, Uttar Banga Krishi Viswavidyalaya.
- Sabesan, T., Saravanan, K. and Satheeshkumar, P. 2016. Studies on heterosis, inbreeding depression and residual heterosis for fruit yield and its components in okra [*A b e l m o s c h u s esculentus* (L.) Moench.]. *Plant Arch.*, **16**(2): 669-674.
- Statista. 2022. https://www.statista.com
- Turner, J.H. 1953. A study of heterosis in upland cotton, combining ability and inbreeding effects. J. Agron., 43: 478-490. [Cross Ref]
- Varma, D.D., Jivani, L.L., Purohit, V.L., Vadavia, A.T. and Rathod, R.K. 2020. Heterosis and inbreeding depression for fruit yield and its component characters in brinjal (*Solanum melongena* L.). *Electronic Journal of Plant Breeding*, **11**(4): 1143-1147. [Cross Ref]