

Research Article**Genotype x environment interactions in rainfed grown greengram (*Vigna radiata*)****K. Anandhi¹, G. Anand² and S. Juliet Hepziba³**¹Krishi Vigyan Kendra, Agricultural College and Research Institute, Madurai. Tamil Nadu Agricultural University²Department of Plant Breeding and Genetics, Agricultural College and Research Institute, Madurai Tamil Nadu Agricultural University³Professor and Head, Agricultural Research Station, Vaigai Dam, Tamil Nadu Agricultural University**E-Mail:**anandhi.k@tnau.ac.in

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Abstract

The low productivity is mainly due to fact that green gram is grown under rainfed environments with vagaries of monsoon. Hence stability of green gram genotypes was analysed during three years in rainfed condition in this study to identify stable genotypes. Among the 15 genotypes along with three checks studied the genotypes G14, G5 and G7 were stable genotypes with high mean yield. These three genotypes show stability for number of pods per plant. The genotype G14 also recorded the lowest value for days to 50 percent flowering with high stability showing early maturity.

Key words

Greengram, Stability, AMMI analysis

Introduction

Green gram [*Vigna radiata* (L) Wilczek] is an important pulse crop rich in protein (24-26%), carbohydrate (54-56%) minerals (4%) and vitamin (3%) required to the tune of 70-80g per capita per day for balance diet of an adult person (Anonymous, 2004). It is an important pulse crop in developing countries of Asia, Africa and Latin America where it is consumed as dry seeds, fresh green pods (Karuppanapandian *et al.*, 2006). More than 70 per cent of total world greengram production is from India (Anonymous, 2016). In India the total production of green gram is 1.71m tons from an area of 3.43m ha with a productivity of 4.98 qha⁻¹ (Anonymous, 2012). Pulse crop act an important role in Indian agriculture as they are rich sources of protein and essential oils for predominant vegetarian population of India (Armugam *et al.*, 2010).

The crop is an important short season summer grown grain legume, well suited to smallholder production under adverse climatic conditions and commonly used in Indian cuisine (Vijayalakshmi *et al.*, 2003). Mungbean is produced in tropical and sub-tropical rainfed environments with little or no impounding of water, and it is prone to drought when soil moisture or rainfall is inadequate to meet plant requirements. The average productivity of greengram over globe is 577 kg ha⁻¹ and in India it is 426 kg ha⁻¹, when compared to the productivity levels over globe, it is far below in India (Anonymous, 2010). The low productivity is mainly due to fact that green gram is grown under

rainfed environments with vagaries of monsoon. Hence stability of green gram genotypes was analysed during three years in rainfed condition in this study to identify stable genotypes.

Materials and Methods

The experiment was conducted during the north east monsoon season at Aruppukottai, Virudhunagar district in Tamil Nadu, India during 2010-13 over three years. The test environment chosen represents typical rainfed which receives less than 300mm rainfall during cropping period. The green gram genotypes used in this study comprised three genetically distinct approved varieties (VBN 2, VBN 3 and CO 7), 15 promising advance lines (Table 1). Six rows in a plot of size 4 x 1.8m were planted with a spacing of 30 cm between rows. Each row contains 40 plants spaced at 10 cm apart.

Additive main effect and multiplicative interaction (AMMI) model was used to determine the stability of the genotypes across environments. The AMMI model first fits the additive effects for the genotypes and the growing environments and multiplicative term for genotype x environmental interactions. The AMMI model according to Farshadfar *et al.* (2011) is presented as

$$Y_{ij} = \mu + g_i + e_j + \sum_{k=1}^n \lambda_k \alpha_{ik} \gamma_{jk} + e_{ij},$$

where Y_{ij} is the yield of the i^{th} genotype in the j^{th} environment, μ is the mean of the i^{th}

genotype minus the grandmean, λ_k is the square root of the eigenvalue of the PCA axis k , α_{ik} and γ_{jk} are the principal component scores for PCA axis k of the i^{th} genotype and the j^{th} environment, respectively, and e_{ij} is the residual. The environment and genotypic PCAscores are expressed as unit vector times the square root of λ_k , i.e., environment PCA score = $\lambda_k^{0.5} \gamma_{jk}$; 0.5; genotype PCA score = $\lambda_k^{0.5} \alpha_{ik}$.

AMMI stability value (ASV) was calculated for each genotype according to the relative contributions of the principal component axis scores (IPCA1 and IPCA2) to the interaction sum of squares. The AMMI stability value (ASV) as described by Purchase *et al.* (2000) was calculated as ASV equal to $\text{IPCA1 Sum of squares} / \text{IPCA2 Sum of squares}$ is the weight given to the IPCA1-value by dividing the IPCA1 sum of squares (from the AMMI analysis of variance table) by the IPCA2 sum of squares. The larger the IPCA score is, either negative or positive, the more adapted a genotype is to a certain environment. Smaller ASV scores indicate a more stable genotype across environments Farshadfar *et al.* (2011).

Results and Discussion

The result of combined analysis of variance (Table 2) showed high significant differences for genotypes, environment and genotype x environment interaction indicating the effect of environment in the genotype x environment interaction, genetic variability and possibility of selection for stable genotypes. As it is indicated by different scientists (Farshadfar and Sutka (2003), Farshadfar and Sutka (2006), when GE interaction was significant, it is possible to proceed and calculate the stability for the tested genotypes.

In AMMI model, principal component analysis is based on the matrix of deviation from additivity or residual will be analyzed. In this respect both the results of AMMI analysis, the genotypes and environment will be grouped based on their similar responses (Pourdad and Mohammadi 2008). Using ANOVA yield sum of square was partitioned into genotype, environment, and GE interaction. GE interaction was further portioned by principal component analysis. According to Farshadfar *et al.* (2011). Stability analysis methods are often used by breeders to identify genotypes that have stable performance and respond positively to improvements in environmental conditions.

AMMI stability value (ASV) indicates the stability of genotypes. It is the distance from zero in a two dimensional scattergram of IPCA1 scores against IPCA2 scores. Since the IPCA1 score

contributes more to the GE sum of square, it has to be weighted by the proportional difference between IPCA1 and IPCA2 scores to compensate for the relative contribution of IPCA1 and IPCA2 total GE interaction sum squares (Fig 1-3b). The distance from zero is then determined using the theorem of Pythagoras (Purchase *et al.*, 2000). Genotypes having low ASV are considered more stable whilst those with high values are less stable genotypes (Hagos and Abay 2013). Genotypes G14, G5 and G7 were stable genotypes with high mean yield. These three genotypes show stability for number of pods per plant. The genotype G14 also recorded the lowest value for days to 50 percent flowering with high stability showing early maturity.

Stability alone for yield performance does not warrant selection since a consistently low yielding genotype can still be stable (Yan and Tinker 2006). In this study, G1, G2, G8 and G9 are low yielding stable genotypes. In some cases the most stable genotypes do not always have the best yield performance (Oliveira and Godoy 2006). The highest yield was recorded by G12 but is not stable.

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References

- Anonymous. 2004. Recommended dietary allowance for Indians. Survey of India Agriculture, Pub. *The Hindu*, **54**.
- Anonymous. 2010. Ministry of Agriculture. Selected State-wise area, production and productivity of green gram, Govt. of India. In: <http://www.indiastat.com>
- Anonymous. 2012. Project co-ordinator's report AICRP on Mullarp crops, IIPR. Kanpur.
- Anonymous. 2016. <http://www.commoditiescontrol.com/eagrtrade/r/staticpages/index>
- Armugam, P.M., K. Anandhi, and B. Selvi. 2010. Stability analysis of yield in black gram. *Legume Research*, **33**: 70-71.
- Farshadfar E, and Sutka J. 2003. Locating QTLs controlling adaptation in wheat using AMMI model. *Cereal Res Commun* **31**: 249-254.

- FarshadfarE, and Sutka E. 2006. Biplot analysis of genotypeenvironmentinteracting in durum wheat using the AMMI model. *Acta Agron. Hung.* **54**: 459-467
- FarshadfarE, N.Mahmodi, and A. Yaghotipoor. 2011. AMMI stability value and simultaneous estimation of yield and yield stability in bread wheat (*Triticum aestivum* L.). *Australian Journal of Crop Science.* **5**: 1837–1844.
- Hagos, H. G. and Abay, F. 2013. AMMI and GGE biplot analysis of bread wheat genotypes in the northern part of Ethiopia. *Journal of Plant Breeding and Genetics.* **1**: 12–18.
- Karuppanapandian T, T. Karuppudurai, P. B. Sinha, A. H. Haniya, K. Manoharan. 2006. Genetic diversity in green gram [*Vigna radiata* (L.)] landraces analyzed by using random amplified polymorphic DNA (RAPD). *Afr J Biotechnol.* **5**: 1214-1219.
- Oliveira, E. and Godoy, I. 2006. Pod yield stability analysis of runner peanut lines using AMMI. *Crop Breeding and Applied Biotechnology.* **6**: 310–317.
- Pourdad, S. S. and Mohammadi, R. 2008. Use of stability parameters for comparing safflower genotypes in multi-environment trials. *Asian J. Plant Sci.* **7**: 100-104
- Purchase, J. L., H. Hatting and C. S. van Deventer. 2000. Genotype x environment interaction of wheat in South Africa: stability analysis of yield performance. *South African Journal of Plant and Soil.* **17**: 101–107.
- Vijayalakshmi P, S. Amirthaveni, R. P. Devadas, K. Weinberger, S. C. S. Tsou, S. Shanmugasundaram. 2003. Enhanced bioavailability of iron from mung beans and its effects on health of school children. Technical Bulletin No. 30. Shanhua, Tainan, Taiwan 741, Republic of China (ROC). Asian Vegetable Research and Development Center. p 32.
- Yan, W. and N. A. Tinker. 2006. Biplot analysis of multi-environment trial data: Principles and applications. *Canadian Journal of Plant Science.* **86**: 623–645.



Table 1. List of green gram genotypes used in this study

S. No.	Genotype	Parentage	Genotype code
1	AGG 29	VGG 77 x ADT 3	G1
2	AGG 31	CoGG 913 x VGG 112	G2
3	AGG 32	CoGG 912 x VGG 112	G3
4	AGG 34	ADG 2080 x VGG 77	G4
5	AGG 35	ADG 2080 x VGG 112	G5
6	AGG 37	VRM 1 x VGG 112	G6
7	AGG 40	ML 267 x ML 682	G7
8	AGG 41	Pusa bold x ML 682	G8
9	AGG 43	ML 267 x Co 4	G9
10	AGG 44	Pusa bold x ML 267	G10
11	AGG 45	Pusa bold x Co4	G11
12	AGG 47	ADT 3 x VGG 77	G12
13	AGG 09 – 068	Co 6 x ML 267/11/1/1	G13
14	AGG 09 – 072	Co 6 x BDYR 2/1/1/1	G14
15	AGG 09 – 077	Co 3 x Jalagon 2/1	G15
16	VBN 2	Check	G16
17	VBN 3	Check	G17
18	Co 7	Check	G18

Table 2. Analysis of variance for AMMI model of green gram for yield

Source	Degrees of freedom	Sum of Squares	Mean Sum of Squares	Variance Ratio	F pr value
Treatments	53	1030377	19441	39.94	<0.001
Genotypes	17	632910	37230	76.49	<0.001
Environments	2	120582	60291	177.09	<0.001
Block	6	2043	340	0.70	0.6506
Interactions	34	276885	8144	16.73	<0.001
IPCA 1	18	166892	9272	19.05	<0.001
IPCA 2	16	109992	6875	14.12	<0.001
Error	102	49645	487		
Total	161	1082064	6721		

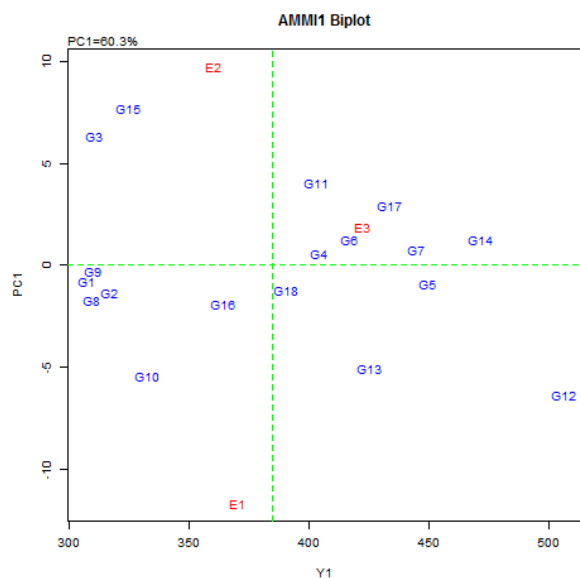


Fig. 1a. Grain yield per plant

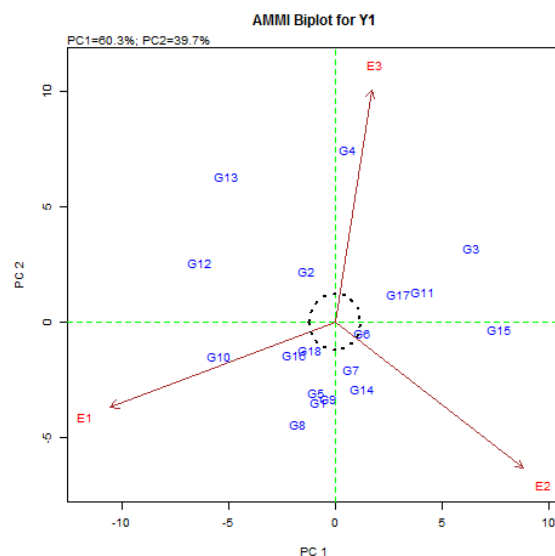


Fig 1b. Grain yield per plant

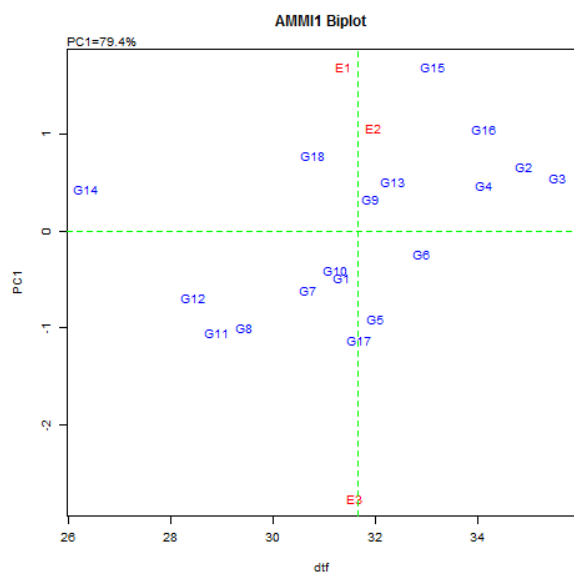


Fig. 2a. Days to 50% flowering

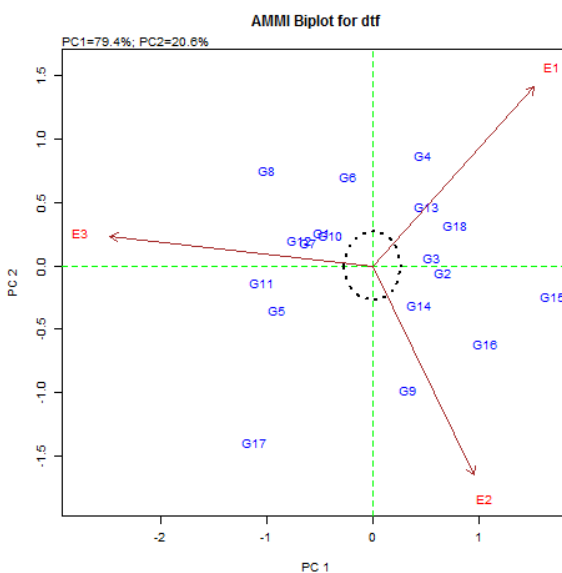


Fig. 2b. Days to 50% flowering

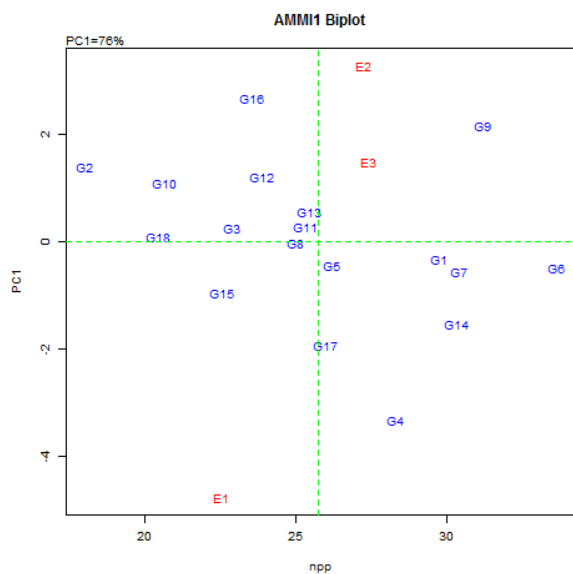


Fig. 3a.Number of pods per plant

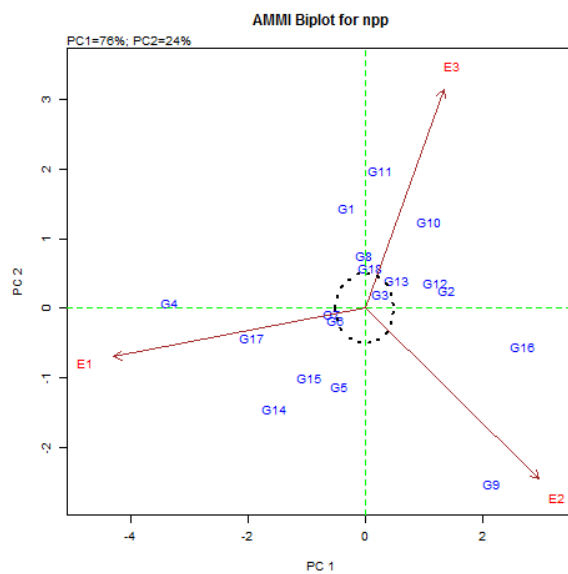


Fig. 3b. Number of pods per plan